



Middle-Upper Devonian palynofloras from Argentina, systematic and correlation

Sol Noetinger ^{a,*}, Mercedes di Pasquo ^b, Daniel Starck ^c

^a Museo Argentino de Ciencias Naturales - CONICET, Angel Gallardo 470, Ciudad A. de Buenos Aires C1405DJR, Argentina

^b Laboratorio de Paleoenestratigrafía y Paleobotánica, CICYTP-CONICET, Dr. Materi & España s/n, Diamante E3105BWA, Entre Ríos, Argentina

^c Tepctrol S.A., Della Paolera 299-Piso 21°, C1001ADA Ciudad A. de Buenos Aires, Argentina

ARTICLE INFO

Article history:

Received 25 April 2018

Received in revised form 19 July 2018

Accepted 29 July 2018

Available online 7 August 2018

Keywords:

Palynology

Taxonomy

Biostratigraphy

Middle-Upper Devonian

Northwestern Argentina

ABSTRACT

This investigation documents the first palynoassemblage spanning the Middle-Upper Devonian from four outcropping sections located in northwestern Argentina. The recovered material includes fairly well-preserved palynomorphs, both from marine and continental origin, including acritarchs, prasinophytes, cryptospores, spores and chlorophytes. In two of the four studied localities chitinozoans were encountered, whilst only one of them have yielded scolecodonts. Five new species are described including one acritarch specimen: *Gorgonisphaeridium impexus* sp. nov., two prasinophytes: *Cymatosphaera robusta* sp. nov. and *Pterospermella simplex* sp. nov., one chitinozoan: *Angochitina plicata* sp. nov. and one miospore species: *Apiculatasporites ruptus* sp. nov. Hierarchical cluster analysis recognize two well-differentiated palynofloras in Argentina for the Early and Middle-Late Devonian. A poor correlation with known Devonian biostratigraphical schemes from Gondwanan and peri Gondwanan regions, support the theory of an endemic flora in northwestern and eastern Gondwana even during the Middle and Late Devonian. The palynoassemblage studied here provides an important source of reference to improve former strata correlation in the Tarija and neighbouring basins.

© 2018 Elsevier B.V. All rights reserved.

1. Introduction

Devonian sediments throughout Argentina are mostly centred in western and northwestern regions (Fig. 1A, B). They have undergone several geological processes that ended up modifying the organic content, resulting in high thermal maturity and the consequential poor recovery of organic material. However, there are several Devonian sections that have yielded palynological assemblages spanning the Lower to Upper Devonian. They comprise material from different sources such as outcrops and exploration boreholes including cuttings and core samples. In western Argentina, the studies include the Talacasto (Le Hérissé et al., 1996; García-Muro et al., 2014); Chigua (Amenábar et al., 2006, 2007), Villavicencio (Rubinstein and Steemans, 2007) and Codo formations (Amenábar and di Pasquo, 2008) (Fig. 2). In northwestern Argentina (NOA), Devonian deposits crop out in the eastern part of the Eastern Cordillera (*Cordillera Oriental*) and the Subandean Ranges, extending also in the subsurface of both, the latter and the Chaco-Salteño Plain (Starck et al., 1993). These deposits are included in the Tarija Basin, shared with Bolivia. Palynological studies in these

regions are for the most part based on subsurface samples (Barreda 1986; Volkheimer et al., 1986; Ottone, 1996; Grahn and Gutiérrez, 2001; Noetinger, 2010, 2015; Noetinger and Di Pasquo, 2011), product of oil exploration surveys of the renowned source rocks from the producing basins of Bolivia and northwestern Argentina (Illrich et al., 1981). These units include the top of the Copo, Caburé, Rincón and Tonono formations in the Chaco-Salteño plain and the Los Monos Formation in Eastern Cordillera (Fig. 2). There is also a small exposure of the Ventana Group (Fig. 2) in the Buenos Aires province, eastern Argentina, which have yielded lycophytes, but no palynomorphs were reported (Cingolani et al., 2003).

Palynostratigraphical schemes based only in ditch cutting samples have limitations. Caving and mud contamination can affect the distribution of elements within a column, resulting in inaccurate palynomorph ranges. Therefore, the use of controls such as subsurface cores and outcrop samples are essential. The Pescado and Piedras formations, exposed in the Eastern Cordillera (Fig. 2), have yielded poor Early Devonian assemblages (Noetinger and Di Pasquo, 2010; Noetinger et al., 2016). Close to the Argentinian-Bolivian boundary there were two studied localities where Early to Middle Devonian palynofloras were recovered (Di Pasquo, 2007; Di Pasquo and Noetinger, 2008). Studied cores spanning the Early Devonian in Noetinger and Di Pasquo (2013) have served as controls in the Chaco-Salteño Plain. Even less numerous are the cases of reliable

* Corresponding author at: Museo Argentino de Ciencias Naturales - CONICET, Angel Gallardo 470, Ciudad A. de Buenos Aires C1405DJR, Argentina.

E-mail addresses: noetinger@macn.gov.ar (S. Noetinger), [\(M. di Pasquo\)](mailto:medipa@cicytp.org.ar), [Daniel.Starck@tecpetrol.com](mailto:D.Starck@tecpetrol.com) (D. Starck).

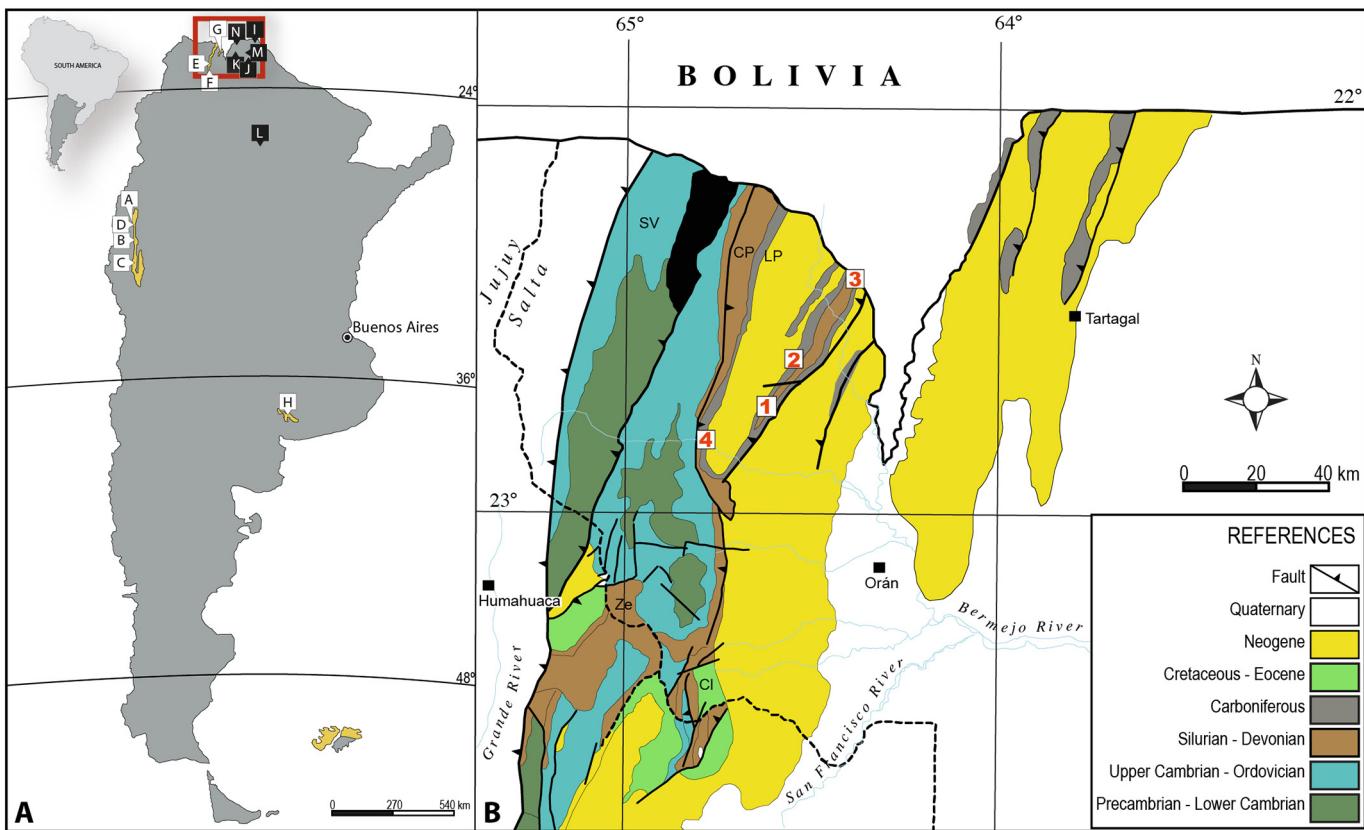


Fig. 1. (A) Devonian outcrops in Argentina and its related studies. Western Argentina: A, Le Hérisse et al. (1996). B, Amenábar et al. (2006, 2007). C, Rubinstein and Steemanns (2007). D, García Muro et al. (2014). Northwestern Argentina: E, Noetinger and di Pasquo (2010). F, Noetinger et al. (2016). G, di Pasquo et al. (2015). Eastern Argentina: H, Cingolani et al. (2003). Studies based in subsurface material: I, Barreda (1986), Noetinger (2015). J, Volkheimer et al. (1986), Noetinger and di Pasquo (2013). K, Ottone (1996). L, Grahn and Gutierrez (2001). M, Noetinger (2010). N, Noetinger and di Pasquo (2011). (B) Geologic map of northern Argentina (modified from Kley and Monaldi, 1999) with the locations studied herein: (1) Angosto de San Ignacio, (2) Angosto del Pescado, (3) Balapuca (sections in both margins of the Bermejo river, political boundary between Argentina and Bolivia), (4) Arasayal.

references for the Middle Devonian in the southern portion of the basin, taking into account only few core samples in Ottone (1996) and Noetinger (2010).

This investigation documents the first palynoassemblage spanning the Middle–Upper Devonian from four outcropping sections located in the Argentinian Subandean (Fig. 1B). The recovered material includes

fairly well-preserved palynomorphs, both from marine and continental origin. A taxonomic detail together with the stratigraphical distribution of the species is presented here, providing a well-supported reference section that will help improve former strata correlation in this basin and neighbouring regions. In addition, a correlation analysis with other Devonian palynofloras from Argentina is addressed.

Chronology Basin/Area		TARIJA			PRECORDILLERA		AUSTRAL RANGES
Geographic units		Supersequences (1)	Chaco-Salteño Plain (2)	Eastern Cordillera (2)	Central (3)	Occidental (3,4)	(5)
Period	Stage	My					
DEVONIAN	Frasnian	383.7	Jollín		Punta Negra	?	
	Givetian		Aguaragüe	Tonono		Chigua	
	Eifelian	393.3		Los Monos		Codo	
	Emsian		Las Pavas	Michicola	Puntilla Negra	Villavicencio	
	Pragian			Rincón		Pircas Negras	
	Lochkovian	419.2	Cinco Picachos	Caburé	Talacasto	?	Ventana Group
SILURIAN	Pridoli	423		Copo	Tamborí		
	Ludlow			Baritú	Los Espejos	?	
				Lipeón			

Fig. 2. Chronostratigraphy of Argentine Devonian units. References: (1, 2) Starck et al. (1993), Starck (1999); (3) Baldis and Peralta (1999); (4) Amenábar and di Pasquo (2008); (5) Limarino et al. (1999).

2. Geological setting

The siluro-devonian succession analysed comprises sandstone-dominated packages alternating with finer-grained shales and siltstones arranged in three supersequences: Cinco Picachos, Las Pavas and Aguaragüe (Fig. 2). The Las Pavas Supersequence is composed of five progradational cycles (I–V, Fig. 3). Each cycle is initiated by grey pelites bearing abundant invertebrates, overlaid by sandstones with hummocky structure finalising, upwards into cross-stratification. Each depositional sequence is bounded by a large-scale flooding surface (Starck et al., 1993). The Supersequence includes the Piedras and Pescado formations (Starck et al., 1993). The Pescado Formation involves fine-grained, dark green-grey greywackes and grey, whitish and yellowish micaceous sandstones with abundant cross-stratification structures interbedded with finer silt- and claystones (Cuerda and Baldis, 1971; Antelo Pérez, 1983). The finely laminated dark shales and siltstones with minor sandstone layers interbedded of the Los Monos Formation follow upward and is part of the overlying Aguaragüe Supersequence.

3. Materials and methods

3.1. Studied localities

Four localities were sampled for palynologic studies in the Argentinian portion of the Subandean Ranges. In the case of the Angosto del Pescado and Balapuca, the sections were surveyed and measured with a Jacob's staff (also known as cross-staff, pole with length marks). The Angosto de San Ignacio and Arasayal section were not measured, and the stratigraphic columns (Fig. 3) are schematic.

3.1.1. Angosto de San Ignacio

Corresponds to an isolated outcrop of shales located in the left margin of the Iruya River, where it cuts across the Cinco Picachos Range at Eastern Cordillera–Subandean boundary ($S\ 22^{\circ}\ 45'\ 57''$; $W\ 65^{\circ}\ 12'\ 14''$ Fig. 1B). According to the local geology, this outcrop involves high levels within the Devonian section. Two samples were extracted from this section but only one was palynological productive (Fig. 3).

3.1.2. Angosto del Pescado

The Pescado river cuts a narrow valley across the structure of the Pescado Range, the Angosto del Pescado section ($S\ 22^{\circ}\ 41'.20.$, $W\ 64^{\circ}\ 34.40.$ Fig. 1B), where 20 samples for palynology (Fig. 3), from the Pescado and Los Monos formations, together with well-preserved lycophyte remains and invertebrates, were collected (Di Pasquo et al., 2015). The fauna of this area is known since the beginning of the twentieth century (e.g. Feruglio, 1933) but the recent collection is still under study.

3.1.3. Arasayal

Four samples were studied from this section (Fig. 3), located in the central part of the Pescado Range ($S\ 22^{\circ}38'40.68''$; $W\ 64^{\circ}31'24.55''$. Fig. 1B). The sampled interval corresponds to a finer-grained shales and siltstones section resting above conglomeratic quartzites. This contact seems to be the flooding surface that separates the Las Pavas and Aguaragüe supersequences. According to the lithological correlation, these levels correspond to the basal section of the Pescado Formation.

3.1.4. Balapuca

This section is located where the Bermejo River cuts the Pescado Range, at the Argentinean-Bolivian border ($S\ 22^{\circ}30'3.96''$; $W\ 64^{\circ}27'37.29''$. Fig. 1B). Twenty three samples were studied from this section (Fig. 3). Geological, including palaeontological, descriptions are included in di Pasquo (2005, 2007) and di Pasquo et al. (2009, 2015).

3.2. Methods

Shale and siltstone and fine-grained sandstone levels were sampled from each of the four localities detailed above. The samples from Angosto del Pescado and Balapuca were processed at the Laboratory of Palynostratigraphy and Palaeobotany CICYTTP (CONICET- Entre Ríos - UADER) in Diamante (Entre Ríos province, Argentina). Standard methodology was performed (Traverse, 2007) with the exception of the use of HCl since the samples did not react to it. Productive samples were improved using other treatments (i.e. HF second time, boiled HCl, KOH, ZnCl), then sieved through a $25\ \mu m$ mesh and mounted with glycerine jelly. The other two localities were processed at the Laboratory of Palaeopalynology (Palaeontology Section, Museo de Ciencias Naturales "Bernardino Rivadavia"- MACN-) following conventional methods (Playford, 1977). Organic residues were then sieved through a $20\ \mu m$ mesh and mounted on standard microscope slides with acrylate (Noetinger et al., 2017). Samples were analysed and illustrated with a trinocular transmitted light microscopes Nikon Eclipse E200 and Leica DM500 both bearing video cameras Amuscope 14 MB at the CICYTTP, and Leica DM2500 with Leica DFC290 camera at MACN. Measurements were carried out through the software ImageJ 1.49v. (Schneider et al., 2012). The position of illustrated specimens in the respective slides is given in England-Finder coordinates, and the materials (slides, residues, rocks) are quoted with the CICYTTP-PI acronym (Repository).

A palynological marine index (PMI) was introduced by Helines et al. (1998), with the purpose of detecting changes in the depositional environment in adjacent samples, with the following formula ($PMI = Marine\ Richness\ (Rm)/(Terrestrial\ Richness\ (Rt) + 1) * 100$). Samples without marine palynomorphs would have a $PMI = 0$, low values of PMI are interpreted as indicative of brackish-water influence, and higher PMI values are interpreted as indicative of marine deposition. Considering that the samples contained fairly well-preserved elements, the marine and terrestrial richness were expressed as number of species per sample rather than genera, as stipulated in de Araujo Carvalho et al. (2006). Because the PMI is based on the palynomorph diversity of terrestrial and marine species, it is therefore used as a substitute for the typical terrestrial/marine ratio.

In order to asses the similarity between the different Devonian localities in Argentina a presence/absence matrix (Supplementary material) was constructed following Barreda (1986), Volkheimer et al. (1986), Le Hérisse et al. (1996), Ottone (1996), Grahn and Gutiérrez (2001), Amenábar et al. (2006, 2007), Rubinstein and Steemans (2007), Noetinger (2010, 2015), Noetinger and Di Pasquo (2010, 2011, 2013), García-Muro et al. (2014) and Noetinger et al. (2016). The total data set comprises 16 localities, spanning the Lower to Upper Devonian and, the list of species representing different taxa (e.g., Phytoplankton, spores, algae, chitinozoans) were taken in consideration, accounting a total of 347. Species left in open nomenclature were not contemplated.

Hierarchical cluster analyses were performed using R version 3.0.1 (R development Core Team 2013). Two dissimilarity matrices were calculated through the package vegan 2.3–5 (Oksanen et al., 2016), using two different coefficients recommended for binary data, Jaccard (see Shi, 1993) and Raup-Crick (Raup and Crick, 1979). The latter were then subjected to hierarchical cluster analysis (Q mode), with the R base package. Since their approaches and assumptions are particular, testing two different algorithms (UPMGA and Complete Linkage) was important in order to weigh the robustness of the groups.

4. Results

4.1. Composition of the palynoflora

The palynological composition of the studied localities present fairly well-preserved, both marine and terrestrial elements. The Angosto de San Ignacio locality provided 31 species: 23

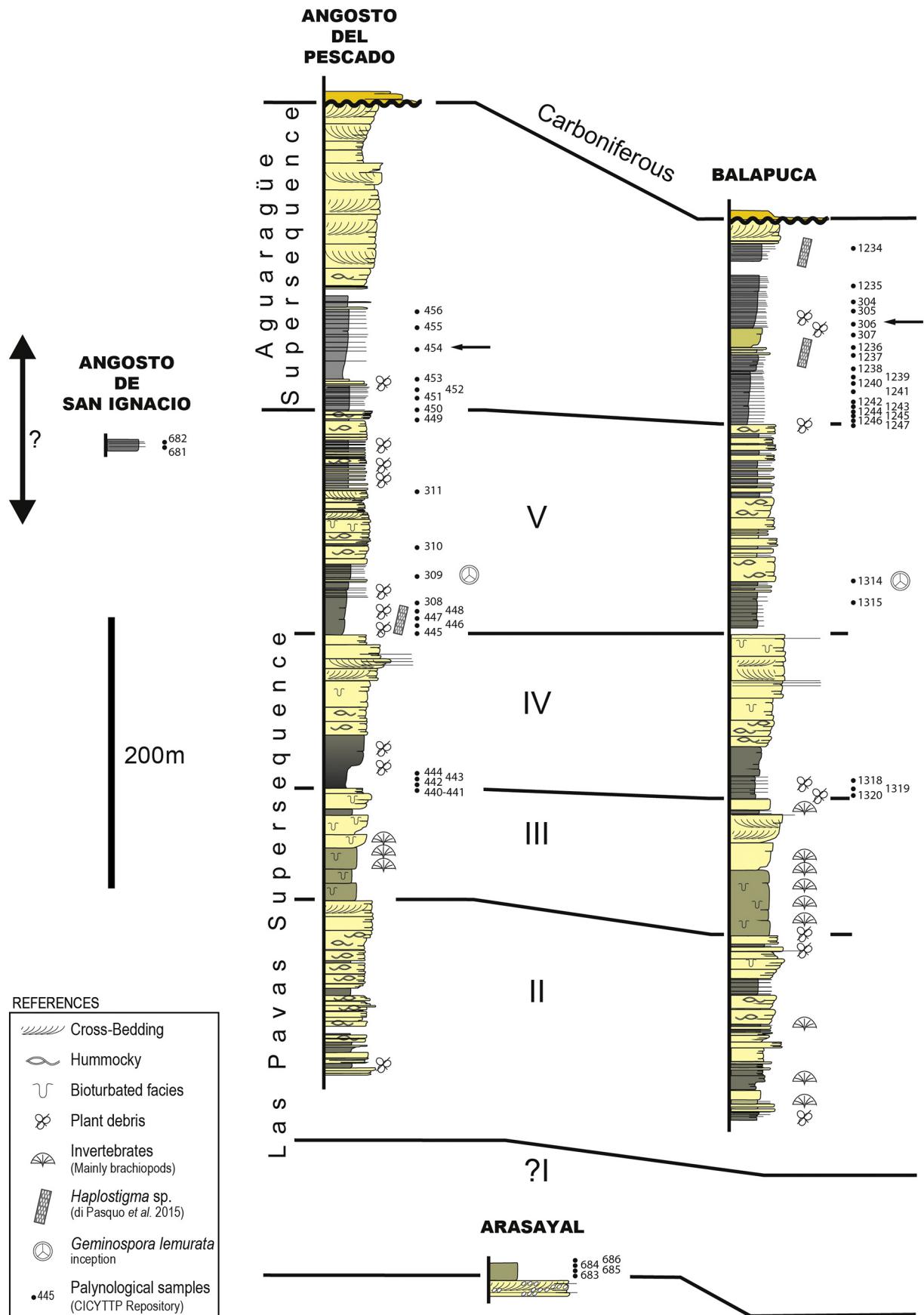


Fig. 3. Composite lithologic logs of the studied sections: Angosto de San Ignacio, Angosto del Pescado, Arasayal and Balapuca with sampling levels. I-V sequences in the Las Pava Supersequence. The arrow marks the transition to the Frasnian in the Angosto del Pescado and Balapuca logs.

representatives of microplankton (15 acritarch and 8 prasinophyte species), 6 spores, 1 cryptospore and 1 chlorophyte species. The Angosto del Pescado locality comprises a very diverse assemblage with different palynological groups, 35 acritarch species, 13 prasinophytes, 8 chitinozoans, 3 chlorophytes, 6 cryptospores and 75 spore species. The Arasayal section yielded a total of 24 species including 12 acritarchs, 6 prasinophytes, 1 cryptospore and 5 spores, and the Balapuca section produced 13 acritarch species, 8 prasinophytes,

4 chitinozoans, 3 chlorophytes, 1 cryptospore, 59 spore species and 3 unidentified scolecodonts (**Plates I–V**).

4.2. Systematic palaeontology

The complete list of recognized taxa are listed by major groups and in alphabetical order in the Appendix. New, left in open nomenclature and species that justify additional remarks are described in the section

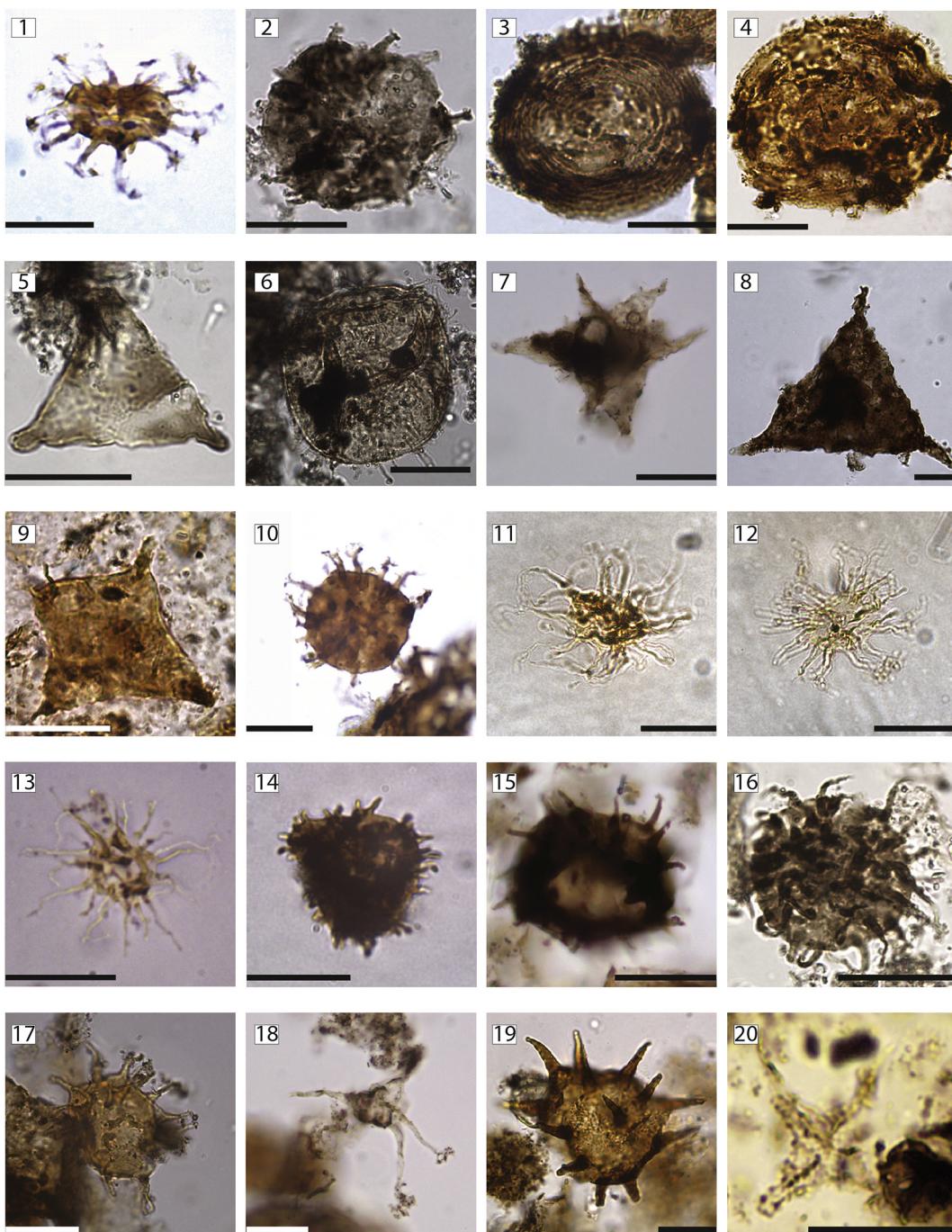


Plate I. Selection of taxa from the studied sections. Each illustration has the sample number and England Finder coordinates specified. Scale = 20 µm. 1. *Ammonidium loriferum* – *Ammonidium garrasinoi* (morphon). CICyTP-PI 682 (2), C55/4. 2. *Baltisphaeridium juliae*. CICyTP-PI 1314(1), K53/1. 3. *Chomotriletes bistchoensis*. CICyTP-PI 454(1), N59. 4. *Chomotriletes vedugensis*. CICyTP-PI 451(1), G28. 5. *Crassiangulina* sp. 1. CICyTP-PI 443(1), X59/3. 6. *Elektoriskos* sp. 1 CICyTP-PI 441(1), B46/2. 7. *Estiastra* sp. cf. *E. culcita*. CICyTP-PI 445(1), C56/4. 8. *Estiastra* sp. cf. *E. rhytidoa*. CICyTP-PI 446(1), P39. 9. *Evittia* sp. cf. *E. sommeri*. CICyTP-PI 308(2), T61. 10. *Gorgonisphaeridium furcillatum*. CICyTP-PI 682(2), F56/4. 11. *Gorgonisphaeridium impexus* sp. nov. Holotype. CICyTP-PI 682(1), K50/2. 12. *Gorgonisphaeridium impexus* sp. nov. Paratype. CICyTP-PI 682(1), D43. 13. *Gorgonisphaeridium impexus* sp. nov. CICyTP-PI 682(2), D45. 14. *Gorgonisphaeridium* sp. cf. *G. absitum*. CICyTP-PI 682(1), Q24/2. 15. *Gorgonisphaeridium* sp. 1. CICyTP-PI 445(1), D56/2. 16. *Gorgonisphaeridium* sp. 2. CICyTP-PI 443(1), N35/3. 17. *Michrystridium flexible*. CICyTP-PI 454(1), Q48. 18. *Ozotobrachion furcillatum*. CICyTP-PI 445(1), B43. 19. Sp. B in Noetinger and di Pasquo, 2011. CICyTP-PI 451(1), T44/2. 20. *Stellinium oppidum* (Deunff) in Quadros 1999. CICyTP-PI 1320(2), D29.

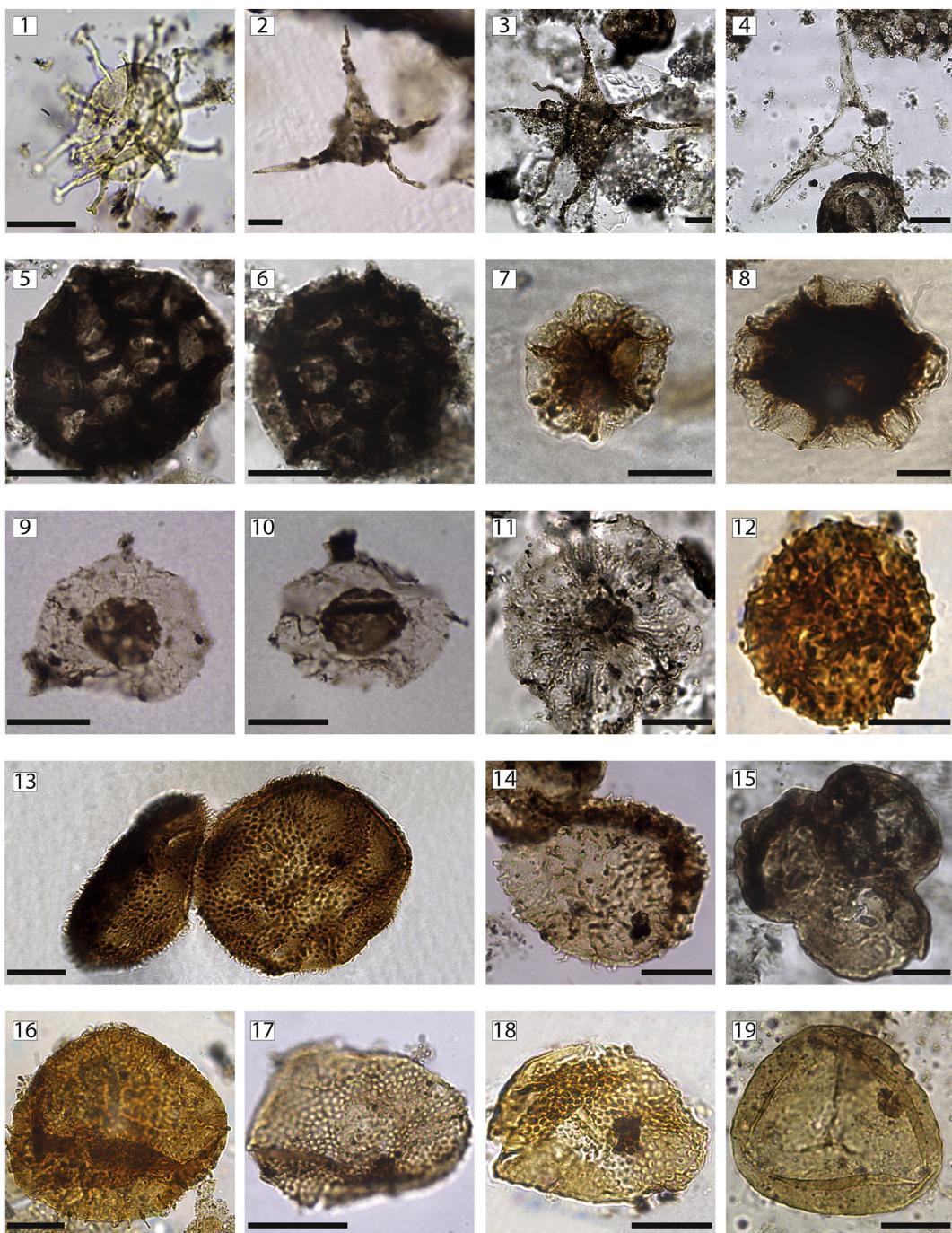


Plate II. Selection of taxa from the studied sections. Each illustration has the sample number and England Finder coordinates specified. Scale = 20 µm. 1. *Umbellasphaeridium* sp. cf. *U. deflandrei*. CICyTP-PI 1239(1), H60/3. 2. *Veryhachium europaeum*. CICyTP-PI 682(2), L29. 3. *Veryhachium pastoris*. CICyTP-PI 444(1), U45/1. 4. *Deflandrastrum?* sp. 1 CICyTP-PI 440 (1), W27/2. 5. *Cymatiosphaera robusta* sp. nov. Holotype. CICyTP-PI 443(1), S46/2. 6. *Cymatiosphaera robusta* sp. nov. Paratype. CICyTP-PI 444(1), E47. 7. *Cymatiosphaera turbinata*. CICyTP-PI 682(1), H36/2. 8. *Duvernaysphaera radiata*. CICyTP-PI 682(1), Q30/4. 9. *Pterospermella simplex* sp. nov. Holotype. CICyTP-PI 684(1), X47. 10. *Pterospermella simplex* sp. nov. Paratype. CICyTP-PI 684(1), O29/1. 11. *Pterospermella* sp. 1. CICyTP-PI 444(1), L38. 12. *Cymbohilates* sp. 1. CICyTP-PI 450(1), U37. 13 *Cymbohilates rubinsteiniae*. CICyTP-PI 682(1), B30/2. 14. *Cymbohilates* sp. 2. CICyTP-PI 451(1), M26/4. 15. *Rimosotetras problematica*. CICyTP-PI 452(1), J38/4. 16. *Anapiculatisporites petilus*. CICyTP-PI 450(1) J38/4. 17. *Apiculatasporites ruptus* sp. nov. Holotype. CICyTP-PI 311(4), D42. 18. *Apiculatasporites ruptus* sp. nov. Paratype. CICyTP-PI 454(1), X34/1. 19. *Archaeozonotriletes* sp. 1. CICyTP-PI 451 (1), X52/2.

below. Species are organized by major groups and following alphabetical order. Morphological terminology related to spores and cryptospores follows the concepts in Punt et al. (2007). The lowest and highest measurements of the spore and cryptospores main axis are detailed, together with the mode in the cases that was possible to calculate. Extra measurements for different variables were also taken in consideration. Stratigraphical important species, new elements and species quoted for the first time in NOA are illustrated.

4.2.1. Systematic descriptions

4.2.1.1. Acritarchs. *Crassiangulina* sp. 1

Plate I, 5

Description: Vesicle triangular in shape with straight or slightly concave sides. Eilyma laevigate. There is one process in each of the vertices of the vesicle. The latter are short homomorphic and are apparently a thickening of the eilyma,

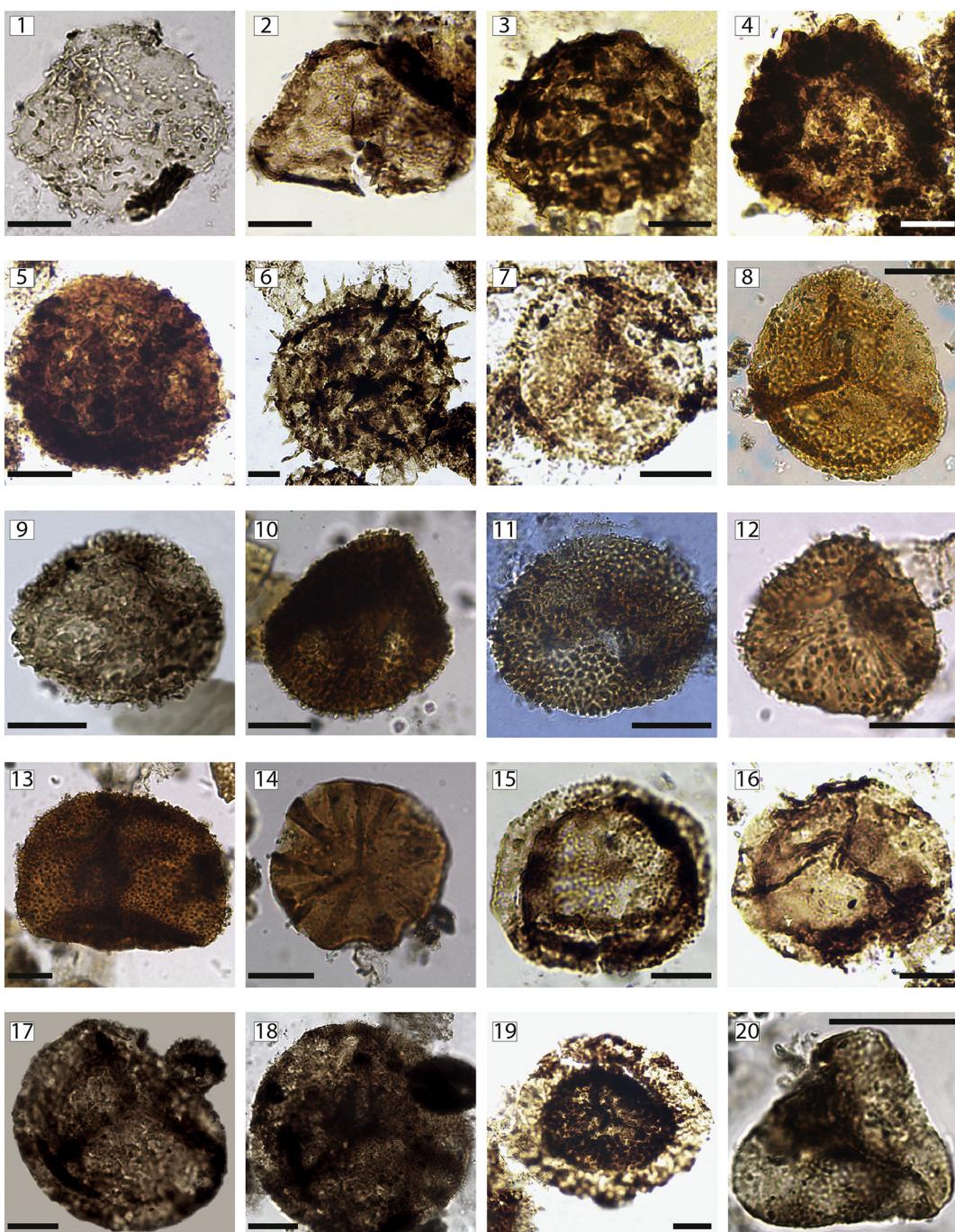


Plate III. Selection of taxa from the studied sections. Each illustration has the sample number and England Finder coordinates specified. Scale = 20 µm. 1. *Biornatispora* sp. cf. *B. dubia*. CICyTP-PI 311(4), D45. 2. *Camarozonotriletes rugulosus*. CICyTP-PI 1239(1), E13. 3. *Chelinospora concinna*. CICyTP-PI 304(2), G44/2. 4. *Chelinospora ligurata*. CICyTP-PI 306(1), H41. 5. *Convolutispora tegula*. CICyTP-PI 1240(1), O45/3. 6. *Corystisporites* sp. cf. *Acanthotriletes horridus* in McGregor and Camfield, 1982. CICyTP-PI 306(2), S24. 7. *Craspedispora paranaensis*. CICyTP-PI 306(2), Z34/3. 8. *Cymbosporites variegatus*. CICyTP-PI 450(1), U55/4. 9. *Dibolisporites* sp. cf. *D. uncatus*. CICyTP-PI 446(1), W51/2. 10. *Dibolisporites* sp. 1. CICyTP-PI 451(1), K47. 11. *Dibolisporites* sp. 1 in Breuer and Steemans, 2013. CICyTP-PI 311(4), D29/4. 12. *Dibolisporites tuberculatus*. CICyTP-PI 454(1), F47. 13. *Dibolisporites turrulatus*. CICyTP-PI 454(1), G55/4. 14. *Emphanisporites robustus*. CICyTP-PI 454(1), N52. 15. *Endoculeospora altobelli*. CICyTP-PI 1245(1), T54/4. 16. *Endosporites longiradiatus*. CICyTP-PI 306(2), E37. 17. *Geminospora antaxios*. CICyTP-PI 443(1), Z31. 18. *Geminospora* sp. 1. CICyTP-PI 444(1), U62/4. 19. *Grandispora megaformis*. CICyTP-PI 307(1), V55. 20. *Granulatisporites concavus*. CICyTP-PI 443(1), W55/2.

Dimensions (one specimen): Overall: 35 µm. Eilyma thickness: 0.9 µm; vesicle side: 27.5–28.2–30.5 µm; process length: 4.8–5.8 µm; process base width: 3.3–4.1 µm.

Occurrence: Angosto del Pescado.

Comparisons: The specimen resembles *Veryhachium improcerum* Wicander and Loeblich 1977 but the latter has shorter processes (3 µm) and is smaller overall (19–20 µm). *Crassiangulina tessellata* Jardiné et al., emend. Wauthoz et al. 2003, has three to eight processes

always verrucate. *C. grotesca* Cramer et al. emend. Wauthoz et al. 2003 bears filamentous elements in each apex and *C. variacornuta* Wauthoz et al. 2003 has complex processes.

Elektoriskos sp. 1

Plate I, fig. 6

Description: Vesicle spherical. Wall thin, laevigate. Numerous stiff, hairlike homomorphic processes (>30) with acuminate or blunt tips that are regularly distributed on the vesicle.

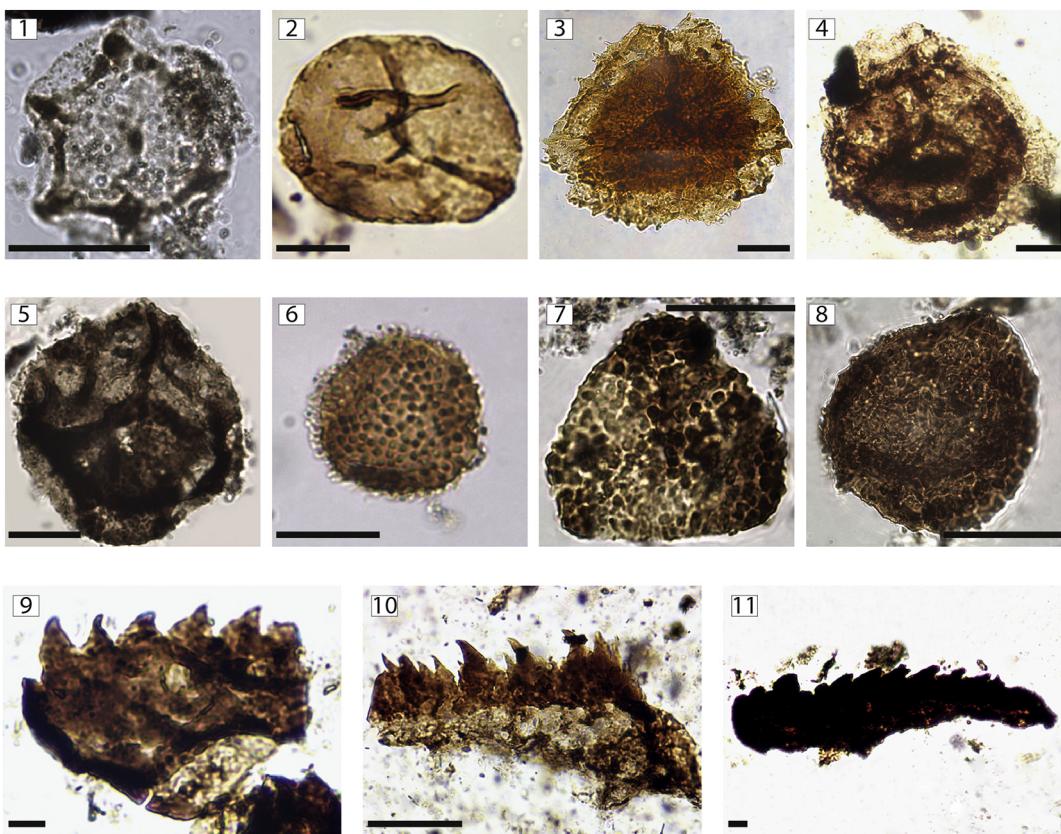


Plate IV. Selection of taxa from the studied sections. Each illustration has the sample number and England Finder coordinates specified. Scale = 20 µm. 1. *Perotritites heterocorus* in Steemans, 1995. CICyTTP-PI 440(1), L58/3. 2. *Punctatisporites piciubaensis*. CICyTTP-PI 1239(1), M48/2. 3. *Samarisporites angulatus*. CICyTTP-PI 450(1), A36/2. 4. *Samarisporites praetervitus*. CICyTTP-PI 1242(1), H27. 5. *Verruciretusispora* sp. 1. CICyTTP-PI 443(1), E41. 6. *Verrucosporites onustus*. CICyTTP-PI 454(1), C29/1. 7. *Verrucosporites* sp. 1 in Breuer and Steemans, 2013. CICyTTP-PI 443(1), U56/4. 8. *Verrucosporites turbulentus*. CICyTTP-PI 456(1), O37. 9–11. Undetermined scolecodonts. (I) CICyTTP-PI 1243(2), N15. (J) CICyTTP-PI 306 (1), T34/4. (K) CICyTTP-PI 1318(3), Y28/2.

Dimensions (3 specimens): Overall: 55–66 µm. Eilyma thickness: 0.7–1.2 µm; vesicle diameter: 40.7–47.9 µm; process length: 2.8–15.3 µm.

Occurrence: Angosto del Pescado

Comparisons: *Elektoriskos* sp. 1 resembles *E. tenuis* Playford in Playford and Dring 1981 but the latter is smaller overall (22–32 µm). *Elektoriskos araiothriches* Loeblich and Wicander 1976 has a smaller vesicle (25 µm) and shorter processes (2–6 µm); *E. intonsus* Loeblich and Wicander 1976 has a more dense distribution of processes and its vesicle is also smaller (31–37 µm); *E. dolos* Wicander and Loeblich 1977 has wider processes at base and the vesicle is smaller (18–21 µm) and *E. villosa* Playford 1981 is smaller overall (21–35 µm) and the eilyma is thicker (1.6–2.6 µm).

Estiastra sp. cf. *E. culcita* Wicander, 1974.

Plate I, 7

Dimensions (one specimen): Overall: 53.6 µm; process length: 17.1–17.7 µm; process base width: 8–14.8 µm.

Occurrence: Angosto del Pescado.

Remarks: The poor preservation prevents to asses the presence of grana in the eilyma.

Estiastra sp. cf. *E. rhytidia* Wicander and Wood, 1981.

Plate I, 8

Description: Vesicle triangular in shape with straight to slightly concave sides. Eilyma laevigate to granulated. There is one process in each of the vertices of the vesicle. The latter are short homomorphic and are apparently a thickening of the eilyma.

Dimensions (3 specimens): Overall: 76.3–103.2 µm. Eilyma thickness: 1.8 µm; vesicle sides: 65.8–72.5–76.3 µm; process length: 6.1–21.3 µm; process base width: 5.9–8.9 µm.

Occurrence: Angosto del Pescado.

Remarks: The poor preservation prevents to asses the presence of grana in the eilyma.

Evittia sp. cf. *E. sommeri* Brito, 1967.

Plate I, 9

Description: Vesicle quadrangular with scabrated eilyma. Short processes at each corner of the vesicle, with barely discernible ramifications.

Dimensions (one specimen): Overall: 44 µm. Process length: 4.4–6.6 µm; Wall thickness: 1.2 µm.

Occurrence: Angosto del Pescado.

Remarks: The poor preservation of the specimen which prevents the clear observation of the bifurcations in the processes and the smaller size avoid the inclusion of the specimen in the species.

Genus: *Gorgonisphaeridium* Staplin, Jansonius and Pocock, 1965.

Type species: *Gorgonisphaeridium winslowii* Staplin, Jansonius and Pocock, 1965.

Gorgonisphaeridium impexus sp. nov.

Plate I, 11–13

Holotype: **Plate I, 11.** Sample CICyTTP-PI 682, K50/2.

Paratype: **Plate I, 12.** Sample CICyTTP-PI 682, D43.

Derivation of the name: The “uncombed” appearance of the processes.

Diagnosis: Vesicle boldly spherical. Wall laevigate under light transmission microscope. Numerous homomorphic processes not less than 15, generally over 30, with rounded bases 1.3–2.1 µm wide, extending 20.8–25.4 µm to end in acuminate recurved tips.

Description of holotype: Vesicle originally spherical 17.2 µm in diameter. Eilyma laevigated, 0.7 µm thick. Numerous homomorphic processes (>20), with rounded bases 1.3–2.1 µm wide, extending 20.8–25.4 µm to end in acuminate recurved tips.

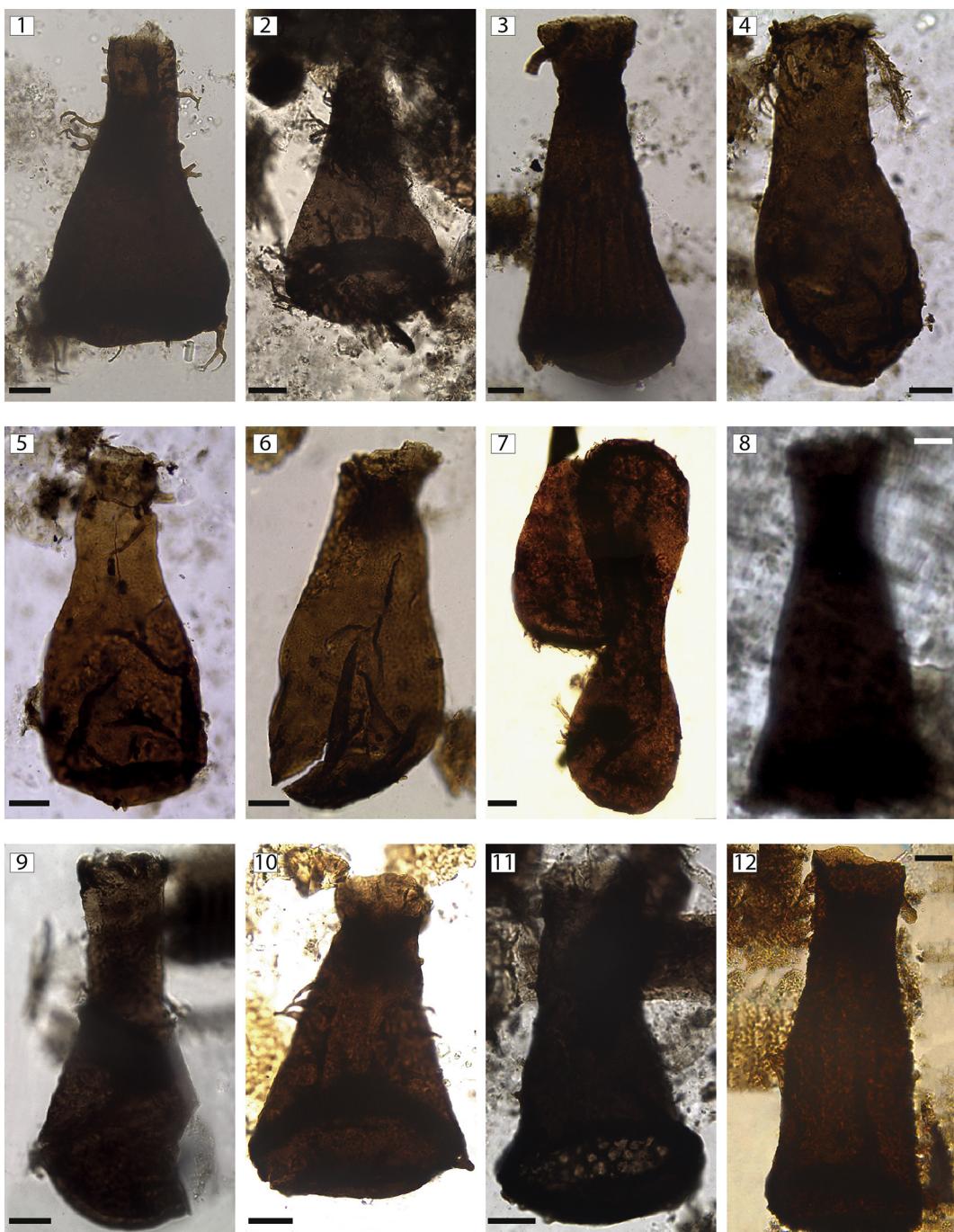


Plate V. Selection of taxa from the studied sections. Each illustration has the sample number and England Finder coordinates specified. Scale = 20 µm. 1–2. *Ancyrochitina morzadeci*. (1) CICyTTP-PI 311(1), X22/2; (2) CICyTTP-PI 311(4), N48. 3. *Ancyrochitina* sp. 1. CICyTTP-PI 451(1), F43. 4–6. *Angochitina plicata* sp. nov. (4) Holotype. CICyTTP-PI 308(2), L43/1; (5) Paratype. CICyTTP-PI 310(2), S39; (6) CICyTTP-PI 310(2), M29/3. 7. *Angochitina* sp. 1. CICyTTP-PI 1314(1), K46/4. 8. *Cyathochitina?* sp. 1 CICyTTP-PI 452(1), Y30/1. 9. *Lagenochitina* sp. cf. *L. praeavelinoi*. CICyTTP-PI 309(3), D52/1. 10. *Ramochitina autasmirimense*. CICyTTP-PI 311(1), Q21/4. 11. *Sagenachitina* sp. 1 CICyTTP-PI 311(4), M46. 12. *Spinachitina* aff. *S. biconstricta*. CICyTTP-PI 450(1), E53/4.

Dimensions (8 specimens): Overall: 32.6–48.5 µm. Eilyma thickness: 0.6–0.7 µm; vesicle diameter: 12–19.5 µm; process basal width: 0.8–2.3 µm; process length: 8.4–25.4 µm.

Occurrence: Angosto de San Ignacio.

Comparisons: *Gorgonisphaeridium absitum* Wicander 1974 has a bigger vesicle (53–66 µm), as well as *G. elongatum* Wicander 1974, which has a bigger vesicle (48 µm average) and an average of longer processes (21 µm). *Gorgonisphaeridium furcillatum* Wicander and Playford 1985 has bi- and trifurcated processes as well as *Gorgonisphaeridium* sp. in Ottone (1996). *Gorgonisphaeridium ohioense* (Winslow) Wicander 1974 has stiffer processes while this specimen bears more flexible and

recurved ones. *Gorgonisphaeridium savertonense* Wicander and Playford 2013 has a wider range for the processes length (10–22 µm).

Gorgonisphaeridium sp. cf. *G. absitum* Wicander, 1974. **Plate I, 14**

Description: Vesicle originally spherical deformed by pyrite precipitation covered by numerous, slightly slender, short processes, rounded at base with simple sharp tips.

Dimensions (one specimen): Vesicle diameter: 28 µm; processes length: 2.7–4 µm; processes basal width: 0.8–1.7 µm.

Occurrence: Angosto de San Ignacio.

Remarks: *Gorgonisphaeridium absitum* is bigger overall (53–66 µm).

Gorgonisphaeridium sp. 1**Plate I, 15**

Description: Vesicle boldly spherical. Wall apparently laeavigate under transmission microscope. Numerous homomorphic processes (>30) with rounded bases that end in acuminate recurved tips that are furcate at the tip.

Dimensions (2 specimens): Overall: 25.6–30 µm. Vesicle diameter: 18.7–20.3 µm; process basal width: 1.7 µm; process length: 3.6–4.6 µm.

Occurrence: Angosto del Pescado.

Comparisons: This specimen resembles *Gorgonisphaeridium* sp. 3 in Barreda (1986) and *Gorgonisphaeridium* sp. in Ottone (1996) but those species are considerably bigger than this one (53 and 61 µm respectively).

Gorgonisphaeridium sp. 2.**Plate I, 16**

Description: Vesicle originally spherical. Eilyma laeavigated. Numerous homomorphic processes (>20), with rounded bases, extending shortly to end in acuminate recurved tips.

Dimensions (5 specimens): Overall: 32.7–46.70 µm. Vesicle diameter: 20.3–32.7 µm; process basal width: 1.3–2.4 µm; process length: 4.6–12.3 µm.

Occurrence: Angosto del Pescado, Arasayal.

Comparisons: *Gorgonisphaeridium* sp. 2 resembles *Gorgonisphaeridium* sp. in Ottone (1996) but the latter has bi- or trifurcate extremities as well as *G. furcillatum*. Wicander and Playford 1985. *Gorgonisphaeridium ohioense* (Winslow) Wicander 1974 has stiffer process that may be flexible from the midpoint to the tip, whilst *Gorgonisphaeridium* sp. 2 has more flexible ones.

Umbellasphaeridium sp. cf. *U. deflandrei* (Moreau-Benoît) Jardiné, Combaz, Magloire, Peniguel and Vachey, 1972.

Plate II, 1

Description: Vesicle sub spherical covered by numerous long processes, rounded at base with simple expanded tips in the shape of funnels.

Dimensions (one specimen): Overall: 64 µm. Vesicle diameter: 34 µm; processes length: 15–18 µm; processes basal width: 1.9–2.6 µm.

Occurrence: Balapuca.

Remarks: *Umbellasphaeridium deflandrei* has shorter processes.

Chlorophycean algae

Genus: *Cymatiosphaera* Wetzel, ex Deflandre, 1954.

Type species: *Cymatiosphaera radiata* Wetzel, 1933.

Cymatiosphaera robusta sp. nov.

Plate II, 5, 6

Holotype: Plate II, 5. Sample CICYTTP-PI 443, S46/2.

Paratype: Plate II, 6. Sample CICYTTP-PI 444, E47.

Derivation of the name: The nature of solid muri.

Diagnosis: Vesicle circular to subcircular with its surface divided into several hexagonal to polygonal fields, which diagonal varies from 6.8 to 19.4 µm long, defined by mostly laeavigated muri 1.2–4.7 µm wide and up to 3.2 µm high that in some cases are surmounted by crests. Some specimens show a larger lacuna up to 24.5 µm wide. Field surfaces often granulated.

Description of holotype: Vesicle circular to subcircular with its surface divided into several hexagonal fields, which diagonal varies from 9.4 to 13.1 µm long. Muri 2.1–3 µm wide. Field surfaces laeavigated. Overall equatorial diameter, 50 µm.

Dimensions (10 specimens): Overall: 40–65 µm (mode: 52 µm).

Occurrence: Angosto del Pescado, Arasayal, Balapuca.

Comparisons: *Cymatiosphaera apia* Ottone 1996 has pentagonal areas and very thin muri. *Cymatiosphaera parvicarina* Wicander 1974 has a prominent a reticulocristate surface. *Cymatiosphaera ambotrocha* Wicander and Loeblich 1977 has a granulated surface.

Deflandrastrum? sp. 1

Plate II, 4

Description: Colony? composed of 4 units. The latter have pyramidal shape with triangular bases. A long and slender process emerge from each vertex of the base, connecting the units between each other.

Dimensions (one specimen): Diagonal -between to the vertices of two units-: 108 µm. Length of unit: 43.7–45 µm; base of unit: 15.1 µm; process length: 18.5 µm.

Occurrence: Angosto del Pescado.

Comparisons: *Deflandrastrum millepedi* Combaz 1962 and *Deflandrastrum* sp. A in Richardson and Ioannides (1973) has a direct contact between the units or through a basal expansion and not by a process. *Deflandrastrum leonardi* Combaz 1962 is similar but the processes that connect the units are wider and shorter.

Genus: *Pterospermella* Eisenack, 1972.

Type species: *Pterospermella aureolata* (Cookson and Eisenack) Eisenack, 1972.

Pterospermella simplex sp. nov.

Plate II, 9, 10

Holotype: Plate II, 9. Sample CICYTTP-PI 684, X47.

Paratype: Plate II, 10. Sample CICYTTP-PI 684, O29/1.

Derivation of the name: The simple nature of the specimens.

Diagnosis: Central vesicle and overall amb spherical. Central body laeavigated, surrounded by an also laeavigated thin equatorial flange, that may fold equatorially resembling a crassitud. The flange does not present radial folds.

Description of holotype: Vesicle circular. Central body circular, laeavigated by an also laeavigated thin equatorial flange without radial folds. Dimensions. Overall: 41.2 µm; central body: 19.1 µm; flange: 9.4–10.2 µm.

Dimensions (12 specimens): Overall: 23–57.5 µm (mode: 41 µm); central body diameter: 12.3–19.1 µm; flange width: 4.5–13 µm. Ratio vesicle – flange: 3.8–1.3 (mode: 1.7).

Occurrence: Angosto del Pescado, Arasayal, Angosto de San Ignacio.

Comparisons: *Pterospermella simplex* sp. nov. resembles *Helios umbelliferum* Pöthe de Baldis 1977, nonetheless as well explained by the author the "lamellae" is supported by regular distributed processes not observed in any of the specimens studied herein. *Pterospermella hermosita* (Cramer) Eisenack et al., 1973 has an ornamented central body; *P. tenellula* Playford 1981 has an ornamented central body and radial folds in the flange. *Pterospermella capitana* Wicander 1974 is bigger (central vesicle 43–62 µm) with a scabrate central body while *P. latibalteus* Wicander 1974 has also a scabrate vesicle and a wider flange (18–26 µm wide) with radial folds.

Remarks: This species dominates the level CICYTTP 684.

Pterospermella sp. 1

Plate II, 11

Description: Central vesicle and overall amb spherical. Central vesicle laeavigated, surrounded by an also laeavigated thin equatorial flange with numerous radial folds.

Dimensions (one specimen): Overall: 57 µm. Vesicle diameter: 17 µm; flange width: 21 µm.

Occurrence: Angosto del Pescado.

Comparisons: *Pterospermella* sp. 1 resembles *P. latibalteus* Wicander 1974 nonetheless the central vesicle of the former seems to be laeavigate. *Pterospermella simplex* sp. nov. is smaller (see above) and it does not present radial folds.

Cryptospores

Cymbophilates sp. 1

Plate II, 12

Description: Cryptospore hilate proximally. Circular to subcircular amb. Distal and subequatorial exospore sculptured with scattered spines, coni and bacula. Elements 1–2.8 µm high and 1.1–2.6 µm wide at base, separated a distance equivalent to one to two base diameters.

Dimensions (3 specimens): Overall: 45.2–50.4 µm.

Occurrence: Angosto del Pescado.

Comparisons: *Cymbophilates* sp. A in Wellman et al., 2000 has only spines as ornamentation. *Cymbophilates* sp. A in Noetinger (2015) has higher elements (2–4 µm high). *Cymbophilates comptulus* Breuer et al., 2007 has smaller elements. (0.5–1.5 µm high) *Cymbophilates* sp. A in

Mehlqvist et al. (2012) resembles *Cymbohilates* sp. 1 but the latter has a denser ornamentation.

Cymbohilates sp. 2

Plate II, 14

Description: Cryptospore hilate proximally. Circular to subcircular amb. Hilum laevigated defined by a subequatorial crassitud. Distal and subequatorial exospore sculptured with scattered slender spines with bulbous bases. Elements 3.6–6.1 μm high and 1.1–2.1 μm wide at base.

Dimensions (2 specimens): Overall: 59.2–66.7 μm .

Occurrence: Angosto del Pescado.

Comparisons: *Cymbohilates* sp. 2 resembles *Cymbohilates* sp. A in Wellman et al. (2000) but the latter has a microgranulated hilum and is smaller overall (35–46 μm). *Cymbohilates* sp. A in Noetinger (2015) has shorter elements (2–4 μm high).

Spores

Genus: *Apiculatasporites* (Ibrahim) emend. Smith and Butterworth, 1967.

Type species: *Apiculatasporites spinulistratus* (Loose) Ibrahim, 1933.

Apiculatasporites ruptus sp. nov.

Plate II, 17,18.

Holotype: Plate II, 17. Sample CICYTP-PI 454, D42.

Paratype: Plate II, 18. Sample CICYTP-PI 311, X34/1.

Derivation of the name: The common preservation state of the specimens.

Diagnosis: Trilete spore. Amb subtriangular to subcircular. Laesurae straight, slightly discernible or covered by lips, extended to the equator. Exine thin laevigated proximally. Distally ornamented with cones 1.2–1.8 μm wide at base and 0.8–1.1 μm high.

Description of holotype: Trilete spore. Amb subcircular, deformed. Laesurae straight, covered by lips 1.4 μm . Exine thin laevigated proximally. Distally ornamented with cones 1.2–1.6 μm wide at base and 1.1 μm high. Overall diameter: 60.4 μm .

Dimensions (16 specimens): 34.4–75 μm .

Occurrence: Angosto de San Ignacio, Angosto del Pescado, Balapuca.

Comparisons: These specimens resemble *Apiculatasporites brevidenticulatus* McGregor and Camfield 1982 but the latter bears more variable ornamentation (grana, coni, spinae). *Apiculatasporites* sp. A in Amenábar et al. (2006) bears higher coni (2–2.3 μm). *Apiculatasporites caperatus* Menéndez and Azcuy 1969 has smaller coni (1 μm high) and grana.

Remarks: The specimens are always plicated and broken, exhibiting the delicate nature of the wall.

Archaeozonotriletes sp. 1

Plate II, 19

Description: Patinated spore, trilete. Amb subtriangular with broad rounded corners to subcircular. Laesurae straight to slightly sinuous extended to the equator, accompanied by labra 1.2–2.2 μm wide. Exine thin proximally, crumpled conforming a concentric fold along the equator. The exine distally is psilate to punctate.

Dimensions (9 specimens): 29–58.6 μm .

Occurrence: Angosto de San Ignacio, Angosto del Pescado, Balapuca.

Comparisons: *Archaeozonotriletes chulus* (Cramer) Richardson and Lister 1969 has a laevigated exine; *A. variabilis* (Naumova) Allen 1965 has a wide patina that could reach the 30% of the total diameter.

Biornatispora sp. cf. *B. dubia* (McGregor) Steemans, 1989.

Plate III, 1

Dimensions (one specimen): Overall: 69.3 μm . Ornamentation: 1.6–2.4 μm wide at base. Ridges 0.8–1.3 μm wide.

Occurrence: Angosto del Pescado.

Remarks: The specimen is bigger overall in comparison with the size range proposed for the species (21–33 μm), and the ornamentation is barely larger.

Dibolisporites sp. cf. *D. uncatus* (Naumova) McGregor and Camfield, 1982.

Plate III, 9

Dimensions (one specimen). Overall: 53.3 μm . Ornamentation: 1.5–1.7 μm wide at base; 1.6–2.3 μm high.

Occurrence: Angosto del Pescado.

Remarks: The ornamentation is smaller than the size range proposed for the species (3–5 μm at basal width and up to 10 μm long).

Dibolisporites sp. 1

Plate III, 10

Description: Spore, trilete. Amb subcircular. Exine laevigated proximally. Equatorial and distal exine ornamented with varied elements mostly biform, bacula, coni and spines densely distributed. The elements are 1.7–2.8 μm in height and 1.5–2.2 μm wide at base.

Dimensions (3 specimens): Overall: 43.8–65.2 μm .

Occurrence: Angosto del Pescado.

Comparisons: *Dibolisporites bullatus* (Allen) Riegel 1973 is much bigger overall (average 137 μm). The sculpture on *Dibolisporites farraginis* McGregor and Camfield, 1982 can show a wide range in distribution (from densely to 8 μm apart) and the ornamentation is more variable in comparison with *Dibolisporites* sp. 1.

Geminospora sp. 1

Plate III, 18

Description: Camerate spore, trilete. Amb subcircular to subtriangular. Laesurae straight extended to the margin of the intexine, accompanied by labra 2.2–3.5 μm wide. Exoexine 3–6 μm thick. *Intexine faintly granular*: Ornamentation distributed closed-packed on the equator and distal face consisting of coni, spines and bacula 1.4–3 μm high, 0.9–2.7 μm wide at base.

Dimensions (5 specimens): 71.6–118 μm .

Occurrence: Angosto del Pescado.

Comparisons: *Geminospora lemurata* Balme 1962 is smaller overall and even though Marshall (1996) proposes a wider size range for this species with specimens up to 110 μm , the ornamentation of *Geminospora* sp. 1 is bigger. *Geminospora antaxios* (Chibrikova) Owens 1971 has grana and microverrucae up to 1 μm high. *Geminospora micropaxila* (Owens) McGregor and Camfield 1982 has shorter elements (usually less than 0.5 μm high). *Geminospora* sp. 1 resembles *Rhabdosporites langii* (Eisenack) Richardson 1965, but the latter has baculi up to 1.5 to 2 μm high.

Verruciretispora sp. 1

Plate IV, 5

Description: Spore, trilete. Amb subcircular. Laesurae straight extended two quarters of the distance to the equatorial margin, accompanied by labra 3.7–4 μm wide. The equatorial margin is darkened due to the density of ornamentation, which define a curvatura perfectae. Contact face laevigated to faintly granulated. Ornamentation distributed on the distal face consists of verrucae that can anastomose forming rugulae and coni.

Dimensions (2 specimens): Overall: 66–79 μm . Ornamentation: 1.6–3.9 μm wide at base.

Occurrence: Angosto del Pescado.

Comparisons: *Verruciretispora dubia* (Eisenack) Richardson and Rasul 1978 has wider verrucae (1.5–10 μm wide at base); *V. ornata* (Menéndez and Póthe de Baldis) Pérez Leyton ex Di Pasquo, 2005 has cylindrical processes of various sizes (1.1–4.4 μm high). These specimens resemble *Verruciretispora pallida* Owens 1971, but the latter has elements that are surmounted by a small cone.

4.2.1.2. Chitinozoans. Ancyrochitina sp. 1

Plate V, 3

Description: Vesicle with a cylindric-conical shape and very short neck that widens towards the aperture. Shoulder absent, flexure discernible. Vesicle laevigated with longitudinal structures. It is observable the development of a process around the aperture being 7.8 μm wide and around the margin.

Dimensions (one specimen): Total length: 203.7 μm , maximum chamber width: 61.5 μm , neck length: 11 μm , neck width: 35.8 μm , aperture: 54.1 μm , flexure angle: 140°.

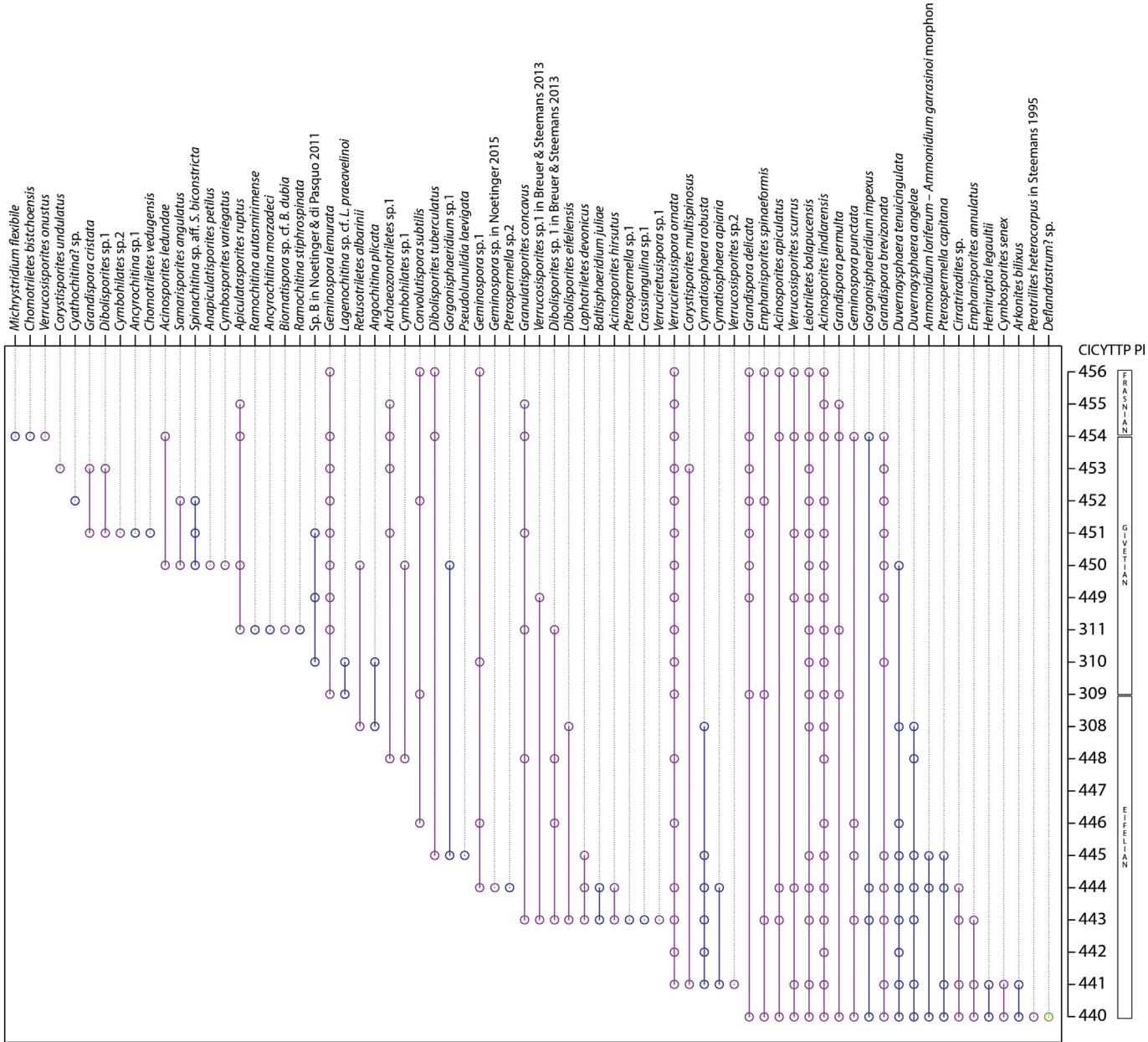


Fig. 4. Stratigraphic distribution of selected species in the Angosto del Pescado section. Marine components (acritarchs, chitinozoans and prasinophytes) are coloured in blue, spores are portrayed in pink and chlorophytes in green.

Occurrence: Angosto del Pescado, Balapuca.

Comparisons: The specimen resembles *Ancyrochitina* sp. A in De Amorim Gaugris and Grahn, (2006) but in the latter the aperture does not evidently expand like in this specimen.

Genus: *Angochitina* Eisenack, 1931.

Type species: *Angochitina echinata* Eisenack, 1931; neotype: *Angochitina echinata* Eisenack, 1964.

Angochitina plicata sp. nov.

Plate V, 4–6

Holotype: Plate V, 4. Sample CICYTTP-PI 310, L43/1.

Paratype: Plate V, 5. Sample CICYTTP-PI 308, S39.

Derivation of the name: The nature of the folded vesicle.

Diagnosis: Vesicle with a cylindric-conical shape, cylindrical neck and rounded base. Shoulder absent, flexure discernible. Vesicle profusely folded mainly laevigated bearing simple and branched spines scattered in the chamber, 6.5–46.8 µm long, 3.7–4.6 µm wide at base and, branched spines around the aperture severely longer 10.6–46.8 µm.

Description of holotype: Vesicle with a cylindric-conical shape, cylindrical neck and rounded base. Shoulder absent, flexure discernible. Vesicle profusely folded mainly laevigated. The aperture is crowned by branched spines 44.5 µm long and 6 µm wide at base. Dimensions: Total length: 170.1 µm, maximum width: 74.8 µm, neck length: 68.5 µm, neck width: 42.5 µm, aperture: 48 µm, flexure angle: 150°.

Dimensions (10 specimens): Total length: 167.8–186.7 µm, maximum width: 72.5–83.5 µm, neck length: 27.5–68.5 µm, neck width: 33.2–42.5 µm, aperture: 30.3–48 µm, flexure angle: 146–154°.

Occurrence: Angosto del Pescado, Balapuca.

Comparisons: *Alpenachitina eisenackii* Dunn and Miller 1964 has neat rows of multi branched processes along the margin, the base of the shoulder and the aperture. This species resembles *Ramochitina boliviensis* Grahn 2002 but the latter bears spongy spines scattered in six rows. The spines surrounding the aperture resemble those of *Alpenachitina crameri* Hutter, 1979, in some of the specimens occurring herein.

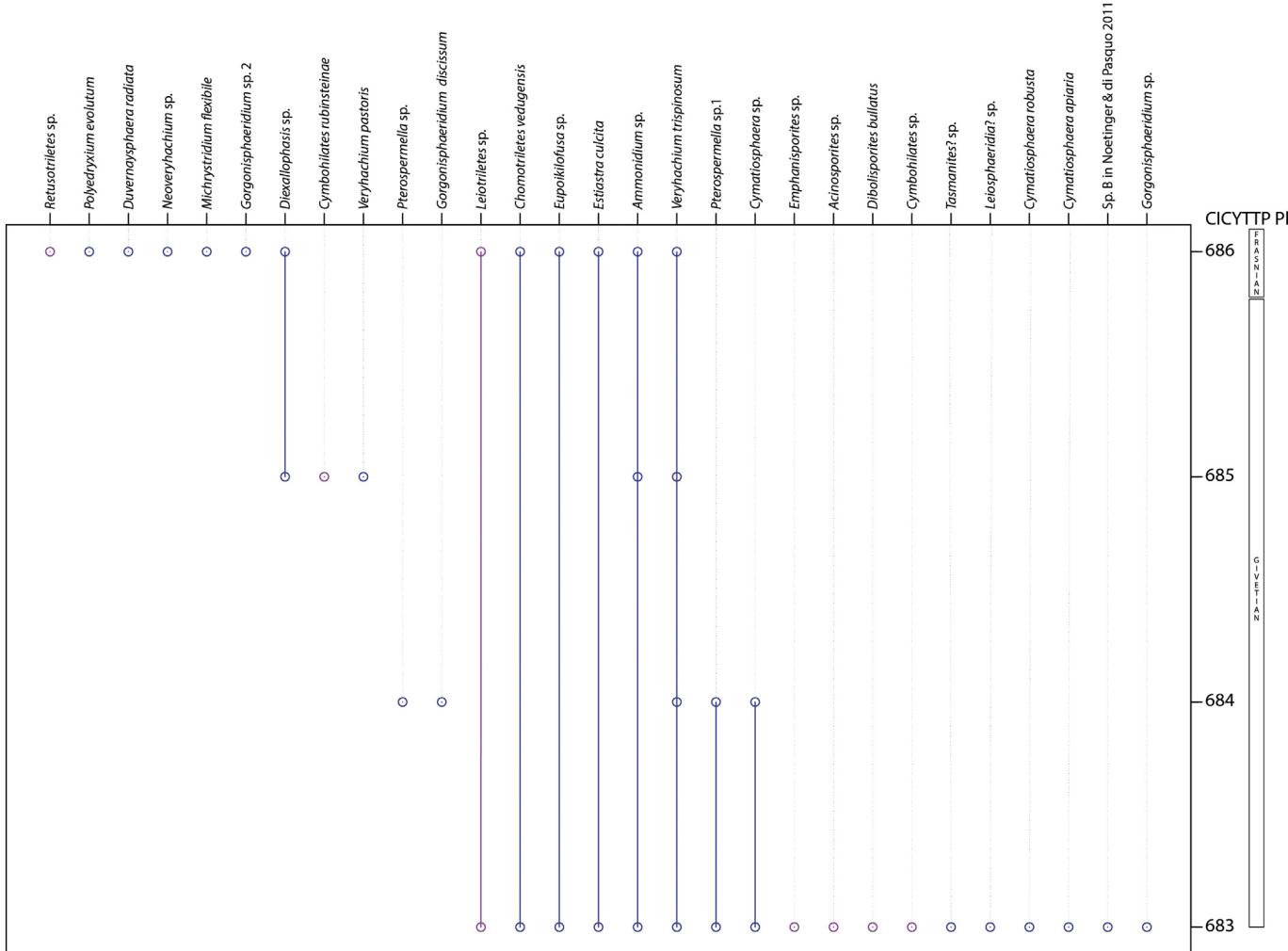


Fig. 5. Stratigraphic distribution of selected species in the Arasayal section. Marine components (acritarchs, chitinozoans and prasinophytes) are coloured in blue, spores are portrayed in pink and chlorophytes in green.

Angochitina sp. 1

Plate V, 7

Description: Vesicle with a cylindric-conical shape, cylindrical neck and rounded base. Shoulder absent, flexure discernible. Vesicle mainly laevigated but with scattered short spines 9–13 µm long, mainly in the margin and stout bifurcated processes in the chamber. The aperture is faintly detected in one of the specimens.

Dimensions (2 specimens): Total length: 187–193.6 µm, maximum width: 77–79.7 µm, neck length: 62.7 µm, neck width: 38.7 µm, aperture: 41.5 µm, flexure angle: 167.6°. Processes: 6.9–19.1 µm long; 3.6 µm wide with rounded bases 1.8–2.8 µm diameter.

Occurrence: Balapuca.

Comparisons: *Angochitina* sp. 1 resembles *A. filosa* Eisenack 1955 but the latter does not have processes. *Angochitina gurupiense* Grahn, Melo & Steemans, 2005 has a neck that widens into a collarette and hair like spines. *Angochitina devonica* Eisenack 1955 has further processes.

Cyathochitina? sp. 1

Plate V, 8

Description: Vesicle with a conical shape, cylindrical neck and rounded base. Flexure discernible.

Dimensions (2 specimens): Overall length: 160–194 µm, chamber maximum width: 73–73.1 µm, margin: 94–102.7 µm, neck length: 28.2–40 µm, neck width: 39–45 µm, aperture: 51.9.1 µm.

Occurrence: Angosto del Pescado, Balapuca.

Remarks: The specimen is assigned to the genus doubtfully due to dubious presence of the carina.

Lagenochitina sp. cf. *L. praeavelinoi* Grahn and de Melo, 2004.

Plate V, 9

Dimensions (2 specimens): Total length: 167.3–178 µm, maximum width: 75–83.6 µm, neck length: 37.3–69 µm, neck width: 31.8–35.2 µm, aperture: 35.9–43.2 µm.

Occurrence: Angosto del Pescado.

Remarks: The specimens encountered herein are smaller than the size range proposed for the species (204–360 µm) and the neck slightly widens towards the aperture.

Sagenachitina sp.

Plate V, 11

Description: Vesicle with a conical shape, cylindrical neck and rounded base. Flexure discernible. Basal margin bearing a perforated carina with lumina in between 3.5 and 4.8 µm diagonal length.

Dimensions (one specimen): Overall length: 149 µm, chamber maximum width: 60.3 µm, neck length: 55.9 µm, neck width: 33.1 µm, aperture: 44.1 µm.

Occurrence: Angosto del Pescado.

Comparison: *Sagenachitna* sp. A in Grahn (2005a) of the Amazonas Basin is considerably bigger (439 µm).

4.3. Age and correlation

The inception and disappearance of several global and South American marker species allowed for a well-constrained age assignment.

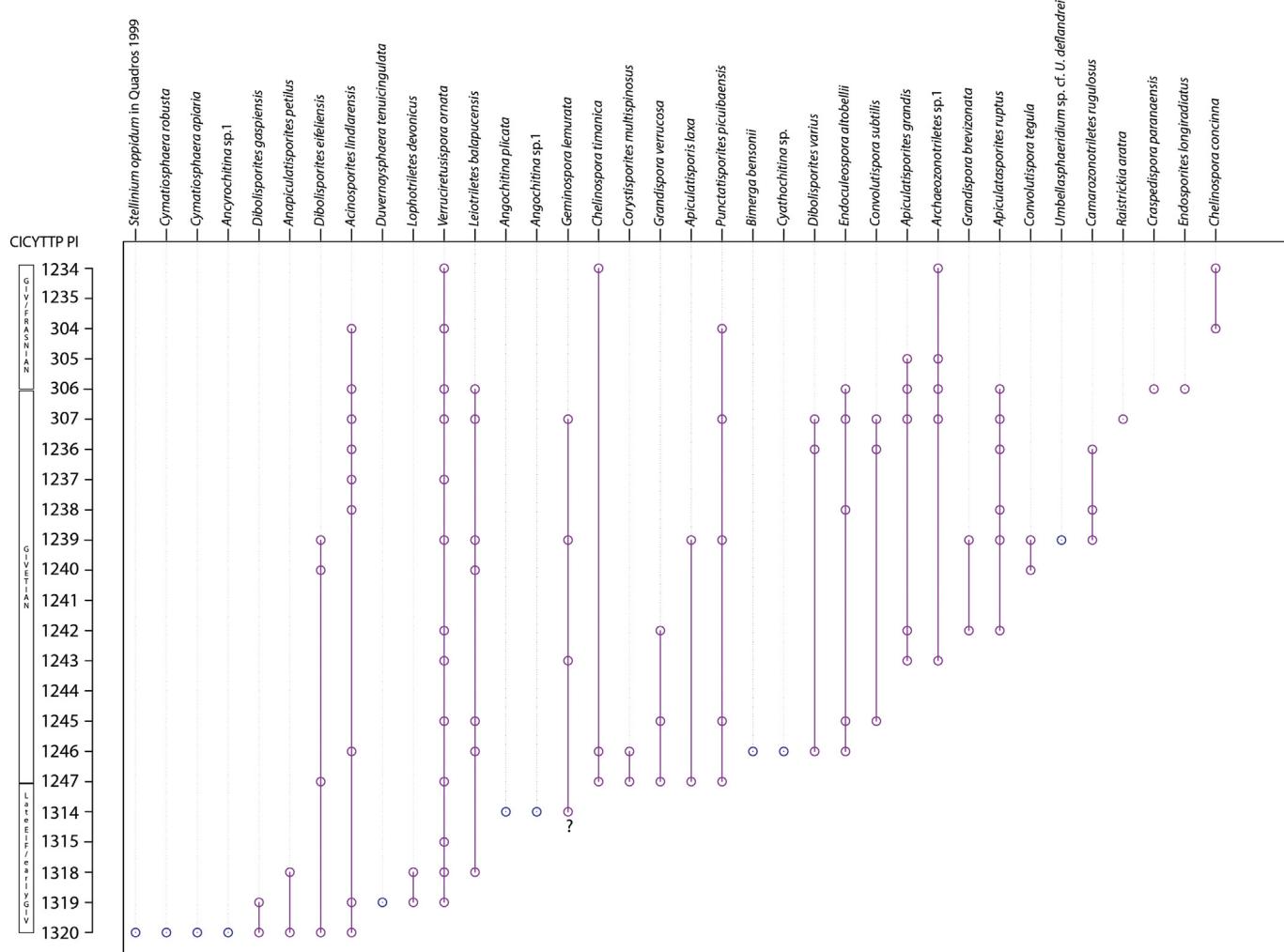


Fig. 6. Stratigraphic distribution of selected species in the Balapuca section. Question mark mean doubtful occurrence. Marine components (acritarchs, chitinozoans and prasinophytes) are coloured in blue, spores are portrayed in pink and chlorophytes in green.

4.3.1. Angosto de San Ignacio

The assemblage recovered from this locality has a very high proportion of marine elements with the input of very few spores (5 species). From the latter *Cymbosporites catillus*, *Dibolisporites turriculosus* and *Retusotriletes simplex* are the only species that have been recorded before. *Cymbosporites catillus* is the only species that is rather more restricted in age than the other two. In southwestern Gondwana, the oldest record of *C. catillus* come from the association SA1 from Argentina, dated late Emsian–mid Eifelian (Noetinger and Di Pasquo, 2011). Also, this species first appears in the highest levels of the Lli and Trg interval zones of the Amazon Basin, of early Givetian age (Melo and Loboziak, 2003) continuing in younger strata. In northwestern Gondwana, in Turkey, *C. catillus* has been quoted as far back in time as the Silurian (Steemans et al., 1996) and it also defines, together with *Samarisporites triangulatus*, the homonymous assemblage zone of middle-upper Givetian age (Breuer and Steemans, 2013). Species from the phytoplankton such as *Cymatiosphaera aparia*, described from Givetian–Frasnian levels in northwestern Argentina (Ottone, 1996), *C. turbinata* known from the Late Devonian of USA and Iran (Wicander and Loeblich, 1977; Ghavidel-Syooki, 1995) and the Givetian of Poland (Turnau and Racki, 1999) and *Gorgonisperidium furcillatum* from the Late Devonian of South America and Euramerica (e.g. Wicander and Playford, 1985; Vavrdová et al., 1996; Gonzalez et al., 2005,

Marynowski et al., 2008; Di Pasquo et al., 2015) allow to bound a Givetian – Frasnian age range for the sample CICYTP PI 682.

4.3.2. Angosto del Pescado

The presence of South American index species (Fig. 4) such as *Leiotriletes balapucensis*, *Grandispora delicata* known to range from the Eifelian (Di Pasquo et al., 2009; Noetinger, 2015) together with, *Acinospores apiculatus*, *Dibolisporites eifeliensis*, *Grandispora permulta*, *Verrucosporites scurrus* and, some species of *Geminospora*, suggest an Eifelian age for the lower part of the section (CICYTP PI 440–309). Microplankton species such as *Arkonites bilixus* and *Duvernaysphaera angelae* support such assignment (see appendix in Di Pasquo et al., 2009; Grahn et al., 2013 and references therein). The inception of *Geminospora lemura* in the level CICYTP PI 309 (Fig. 3) marks the end of the Eifelian and beginning of the Givetian (see Loboziak and Strel, 1995). The disappearance of the chitinozoans *Ancyrochitina morzadeci* and *Spinachitina aff. S. biconstricta* denotes the top of the late early Givetian interval of the Trg Zone recognized in the Amazonas and Paráiba basins (Melo and Loboziak, 2003; Grahn et al., 2006). Finally, the occurrence of *Chomotriletes bistchoensis* and *Michrystridium flexibile* in the level CICYTP PI 454 allow for the appraisal of a Frasnian age for the top of the section. Both species are known from the Frasnian of Euramerica and the former also from Australia and northwestern

Table 1

PMI values, from left to right the localities are organized in north–south direction. Samples are distributed in accordance with the age column on the left, but their separation within each locality are not proportional. Arrow represent the inception of *Geminospora lemurata* (late Eifelian – early Givetian) in both, Balapuca and Angosto del Pescado, localities.

Age	Balapuca		Angosto del Pescado		Angosto de San Ignacio		Arasayal	
	Samples	PMI	Samples	PMI	Samples	PMI	Samples	PMI
Givetian	CICYTTP-PI 1234	4	CICYTTP-PI 456	14			CICYTTP-PI 686	367
	CICYTTP-PI 1235	0	CICYTTP-PI 455	7			CICYTTP-PI 685	200
	CICYTTP-PI 304	0	CICYTTP-PI 454	16			CICYTTP-PI 684	500
	CICYTTP-PI 305	0			CICYTTP-PI 682	314	CICYTTP-PI 683	217
	CICYTTP-PI 306	6						
	CICYTTP-PI 307	0						
	CICYTTP-PI 1236	6						
	CICYTTP-PI 1237	0						
	CICYTTP-PI 1238	11						
	CICYTTP-PI 1239	21						
	CICYTTP-PI 1240	0						
	CICYTTP-PI 1241	0	CICYTTP-PI 453	10				
	CICYTTP-PI 1242	0	CICYTTP-PI 452	15				
	CICYTTP-PI 1243	0	CICYTTP-PI 451	33				
	CICYTTP-PI 1244	18	CICYTTP-PI 450	32				
	CICYTTP-PI 1245	0	CICYTTP-PI 449	20				
	CICYTTP-PI 1246	8	CICYTTP-PI 311	40				
	CICYTTP-PI 1247	18	CICYTTP-PI 310	41				
	CICYTTP-PI 1314	50	CICYTTP-PI 309	50				
	CICYTTP-PI 1315	30	CICYTTP-PI 308	92				
	CICYTTP-PI 1318	63	CICYTTP-PI 448	47				
	CICYTTP-PI 1319	54	CICYTTP-PI 446	44				
	CICYTTP-PI 1320	76	CICYTTP-PI 445	160				
			CICYTTP-PI 444	96				
			CICYTTP-PI 443	70				
			CICYTTP-PI 442	136				
			CICYTTP-PI 441	100				
			CICYTTP-PI 440	122				

Gondwana (Wicander, 1974; Wicander and Playford, 1985; Ghavidel-Syooki, 1995).

4.3.3. Arasayal

The lower section is characterized by the presence of scarce but important Givetian species (Fig. 5) such as *Chomotriletes vedugensis* and *Cymatiosphaera apiaria*. *Dibolisporites bullatus* is recorded in the *Grandispora pseudoreticulata* Zone of Eifelian–?early Givetian age in southwestern Gondwana (Suárez Soruco and Lobo Boneta, 1983; Limachi et al., 1996), in northwestern Gondwana occurs in the *Latosporites ovalis-Dictyotriletes biornatus* Assemblage Zone Breuer and Steemans 2013 while in Euramerica is recorded earlier, in the Givetian–Frasnian boundary (Lobozia and Strel, 1980). The co-occurrence of the species mentioned above place the samples (CICYTTP PI 683–685) in the Givetian. The rest of the species in the assemblage are badly preserved elements left in open nomenclature and species with broad age ranges. The top of the section represented by level CICYTTP PI 686 includes the occurrence of *Michrystridium flexibile* recognized in the Frasnian of Euramerica (see above).

4.3.4. Balapuca

Dibolisporites eifeliensis, *D. echinaceus*, *Acinosporites lindlarensis*, *Apiculiretusispora brandtii* and *A. plicata*, in between other species characterize the *Emphanisporites annulatus-Camarozonotriletes sextantii* Assemblage Zone of early and early late Emsian age in the Old Red Sandstones (Richardson and McGregor, 1986). In this case these species co-occur with other ones (Fig. 6) that have more modern inceptions worldwide such as *Leiotriletes balapucensis* (see above), *Chelinospora timanica* and *Geminospora lemurata*, which are included in the *Geminospora lemurata-Chelinospora ex gr. ligurata* Interval Zone (Lli) in the Amazonas Basin (see appendix in Di Pasquo et al., 2009). *Leiotriletes*

balapucensis is also cited in the *Scylaspora rugulata-Grandispora libyensis* Assemblage Zone in northwestern Gondwana, *G. lemurata* appears further high (*Geminospora lemurata* Interval Zone). *Grandispora verrucosa* is known from the Eifelian of southwestern Gondwana (Menéndez and Pöthe de Baldis, 1967; Di Pasquo, 2007). All of the latter suggests a late Eifelian– early Givetian age for the lower part of the section (CICYTTP PI 1320–1247, Fig. 3). The inception of *Endoculeospora altobelli*, known from the middle Givetian (Di Pasquo, 2007) allows to infer a Givetian s.l. age for the section comprised in between the samples CICYTTP PI 1246–307. A Givetian–Frasnian age for the uppermost of the section (CICYTTP PI 306–1234, Fig. 3) is supported by the presence of *Craspedispora paranaensis* that ranges from the upper Eifelian to Givetian in southwestern Gondwana to Frasnian in northwestern Gondwana (Breuer and Steemans, 2013 and references therein), *Endosporites longiradiatus* quoted for the Givetian–Frasnian of Argentina, Bolivia and Paraguay (Menéndez and Pöthe de Baldis, 1967; Ottone, 1996; Di Pasquo, 2007; Di Pasquo et al., 2009), and *Chelinospora concinna*, which its first occurrence defines the *Ancyrospora langii-Chelinospora concinna* Assemblage Zone in northwestern Gondwana (Breuer and Steemans, 2013).

4.4. Palynological marine index

We calculated the Palynological Marine Index (PMI) in order to analyse the evolution of the depositional environments (Table 1). The progress of the index is better seen in the localities with more samples (i.e. Angosto del Pescado, Balapuca). From north to south, Balapuca seems to have been deposited in mostly a nearshore environment, whilst Angosto del Pescado, farther south and west, transition from a seaward environment, during the Eifelian, to nearshore in the Givetian– Frasnian (Table 1). As described above, both the Las Pavas

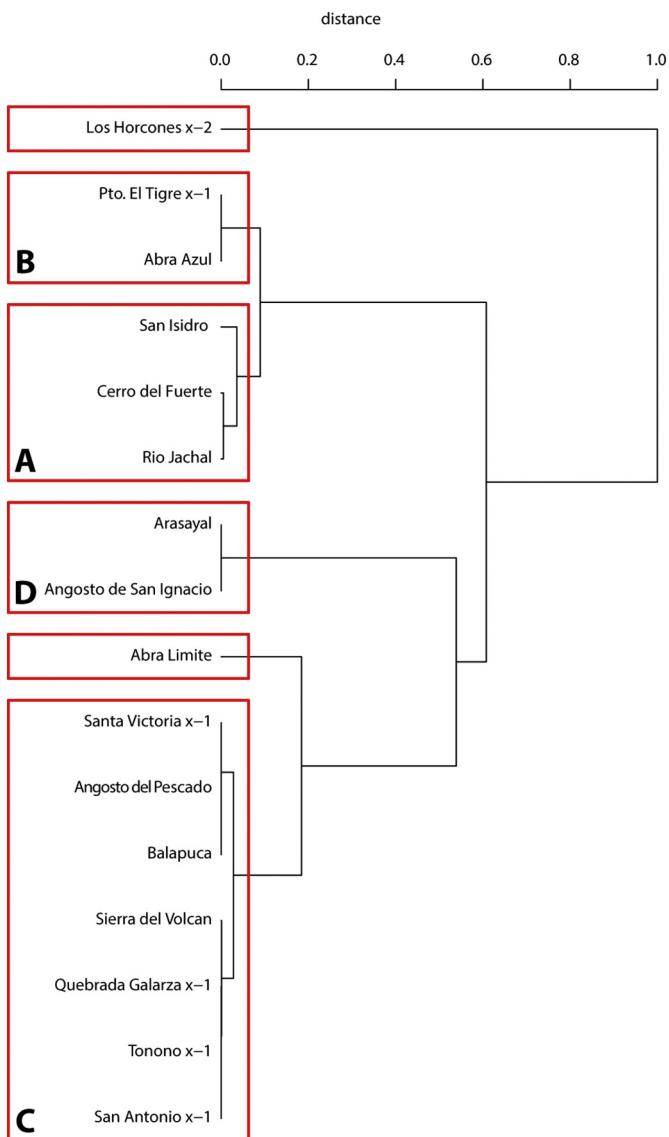


Fig. 7. Dendrogram plot derived from the Raup dissimilarity index using UPGMA algorithm. Group A comprises localities dated as Early Devonian, group B range from late Lochkovian to the Emsian and group C encloses localities assigned to the Middle–Upper Devonian. Group D also dated as Middle–Upper Devonian. References: Los Horcones x-2 Grahn and Gutierrez (2001), Puesto El Tigre x-1 (Volkheimer et al., 1986; Noetinger and di Pasquo, 2013), Abra Azul (Noetinger et al., 2016), San Isidro (Rubinstein and Steemans, 2007), Cerro del Fuerte (Le Hérisse et al., 1996), Río Jáchal (García Muro et al., 2014), Arasayal (this paper), Angosto de San Ignacio (this paper), Abra Límite (Noetinger and di Pasquo, 2010), Santa Victoria x-1 (Barreda, 1986; Noetinger, 2015), Angosto del Pescado (this paper), Balapuca (this paper), Sierra del Volcán (Amenábar et al., 2006, 2007), Quebrada Galarza (Ottone, 1996), Tonono x-1 (Noetinger, 2010) and San Antonio x-1 (Noetinger and di Pasquo, 2011).

and Aguararagüe supersequences, (Figs. 2, 3), are upward – coarsening sequences, a shelf that shallows. Coincidentally, is in these deposits where the chitinozoans were more numerous (Fig. 3). Chitinozoans are known to occur from near-shore to deep oceanic environments. Paris (2006) also links the shallowing of the shelf with the increase of the abundance of this faunule. The latter seems to be the case in the localities studied here (Angosto del Pescado, Balapuca). The disappearance of some particular species (i.e. *Ancyrochitina morzadeci* and *Spinachitina* aff. *S. biconstricta*) could imply the turnover of this fauna, as stated in Grahn (2005b); poor preservation of the organic material due to the deposition of more coarser sediments or both facts combined. Unfortunately the regional Carboniferous erosion (Starck et al., 1993), prevents the recovery of Upper Devonian

material. Towards the south and west, the localities Angosto de San Ignacio and Arasayal, even with their small number of samples, show a very different depositional environment, with PMI values in the order of hundreds (Table 1), implying an offshore depositional environment.

4.5. Hierarchical cluster analysis

This method serves as data visualization better than a statistical analysis per se (Hammer and Harper, 2006) and its application, in this instance, was to explore the existence of any pattern of correlation between the localities and/or basins, during a particular time range. Cluster analysis performed with the two different algorithms and indices (Fig. 7, S. Fig. 1) coincide roughly in the composition of the groups. The following analysis was based in the dendrogram reached with Raup dissimilarity index and UPMGA (Fig. 7) because it is the one that best reflects our observations. The differences between the dendograms might be in the intrinsic characteristics of each algorithm and/or index (i.e. Complete Linkage is biased towards producing clusters of equal diameter, and is also very sensitive to outliers - Milligan 1980, SAS Institute Inc. 1999).

Group A comprises most of the localities from Precordillera (San Isidro, Cerro del Fuerte, Río Jáchal) dated as Early Devonian. Group B includes two localities, one from the Chaco-Salteña plain (Puesto El Tigre x-1) and one from the Subandean (Abra Azul) that range from late Lochkovian to the Emsian. Group C encloses localities assigned to the Middle–Upper Devonian and group D is constituted by the localities Arasayal and Angosto de San Ignacio studied herein, which are also dated as Middle–Upper Devonian. The latter dissociation might be due to differential environmental conditions and richness of the samples. Cluster C has very rich localities while Arasayal and Angosto de San Ignacio are conformed by a lower proportion of species and a higher proportion of marine and brackish elements (Table 1). Los Horcones x-2 and Abra Límite, are isolated probably because the low count of species.

5. Discussion

Few biostratigraphical schemes have been attempted for the southwestern Gondwanan region (e.g. Suárez Soruco and Lobo Boneta, 1983; Limachi et al., 1996; Melo and Loboziak, 2003; Grahn, 2005b) as well for Euramerica and northwestern Gondwana (e.g. Richardson and McGregor, 1986; Strel et al., 1987; Breuer and Steemans, 2013) (Fig. 8). Some are more rigorous than others, including not only illustrations but systematic descriptions. The Argentinian palynological assemblages despite its poor preservation, present a considerable number of global recognized species (see appendix in Di Pasquo et al., 2009) that have served to characterize the stages within the different biozonations (e.g. Noetinger, 2010, 2015; Noetinger et al., 2016; Aráoz et al., 2016 and references therein); but on the other hand, there are several key species used to define those zones that are absent in northern Argentina. There is also a diachronism in the first and last occurrences and acme events of species, i.e. characteristic species from the Givetian *Geminospora lemurata-Rhabdosporites langii* Assemblage Zone (Breuer and Steemans, 2013) such as *Rhabdosporites langii*, *Geminospora lemurata*, *Corystisporites undulatus* and *Dibolisporites turriculatus* co-occur with known younger species in the top of the succession studied here. From the latter, *Chomotrites bistichoensis*, known from the Upper Devonian of Euramerica (Staplin, 1961; Wicander and Playford, 1985) and northwestern Gondwana (Ghavidel-Syooki, 1995), also reworked in the Upper Carboniferous of the Tarija Basin (Di Pasquo, 2002), and *Michrystridium flexible* from the Upper Devonian – Lower Mississippian of Euramerica (Wicander, 1974). There are, also, species with younger records that occur earlier in this assemblage. *Cymatiosphaera apiaria*, described from Givetian-Frasnian strata in the Tarija Basin (Ottone, 1996); *Estiastra culcita*, from the Upper Devonian of Euramerica

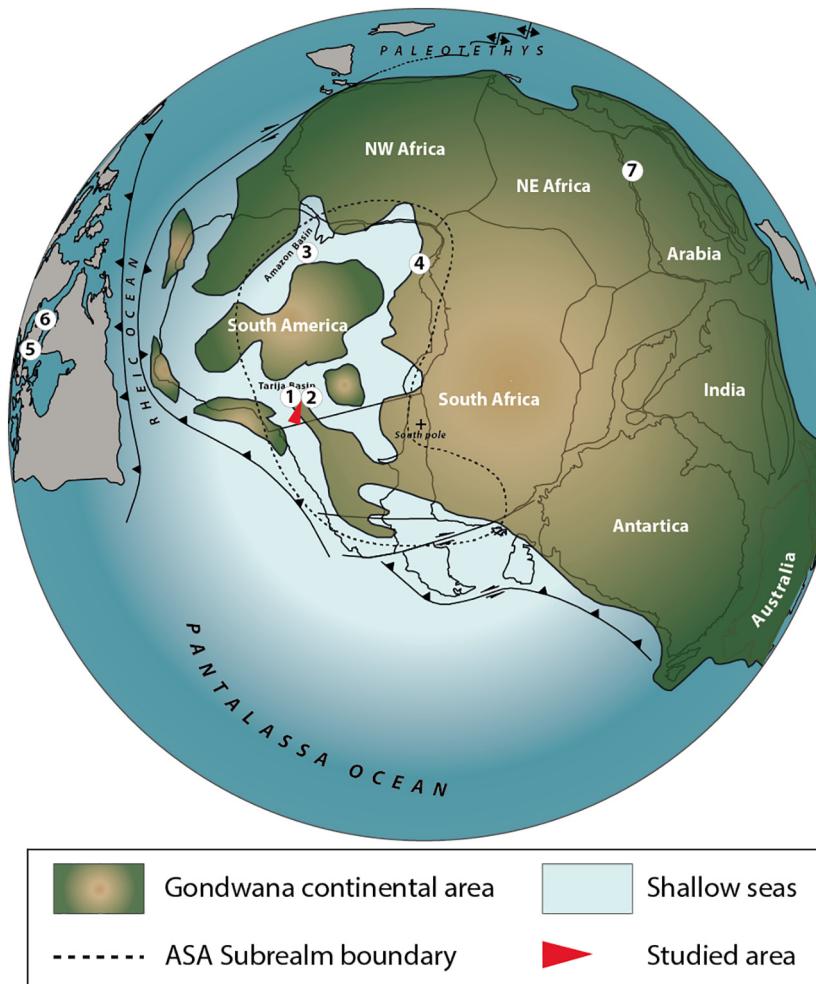


Fig. 8. Palaeogeographical map featuring the Gondwana continent during the Middle Devonian (after Uriz et al., 2016 and references therein) showing the locations where bioestratigraphical schemes, cited in text, were proposed. Southwestern Gondwana. (1) Suárez Soruco and Lobo Boneta (1983). (2) Limachi et al. (1996). (3) Melo and Lobozik (2003). (4) Grahn (2005). Euramérica. (5) Richardson and McGregor (1986). (6) Strel et al. (1987). Northwestern Gondwana. (7) Breuer and Steemans (2013). Dashed line encircles the Afrosouthamerican Subrealm (ASA) (after di Pasquo et al., 2009, 2015).

(Wicander, 1974; Wicander and Playford, 2013), and *Pterospermella capitana* from the Upper Devonian of Euramerica (Wicander, 1974; Martin, 1984; Filipiak, 2005) and the Givetian – Frasnian of the Tarija Basin (Ottone, 1996; Noetinger, 2010). Furthermore, Kemp et al. (1977) suggest to use biozones cautiously for correlation, since the palynoassemblages may be affected by local climatic and palaeoecological factors. The geographical position of the continents (Fig. 8) may have also affected the stratigraphical distribution or time of inception of the species in this region and finally, the interaction of all the latter circumstances. The groups conformed by hierarchical clustering recognized two well-differentiated periods of time for Argentina, Early and Middle-Late Devonian (Fig. 7). Nonetheless, a correlation between the different basins of Argentina was rather inconclusive (e.g. Sierra del Volcán – western Argentina – is grouped with northern localities). Several endemic palynological species closely distributed, both terrestrial and marine (e.g. *Acinosporites ledundae*, *Grandispora cristata*, *G. delicata*, *G. pseudoreticulata*, *Lagenochitina praeavelinoi*, *Spinachitina* aff. *S. biconstricta*) support the singularity of the regional conditions, which altogether conformed the Afrosouthamerican Subrealm (ASA) (Fig. 8, Di Pasquo et al., 2009, 2015). All of the latter reflects the necessity for a local biostratigraphical proposal, which with the addition of more localities would be a plausible goal to achieve. The palynoassemblage studied in this paper provides an invaluable source of reference to improve former strata correlation in the Tarija and neighbouring basins.

Acknowledgments

Mr. Sebastián Mirabelli (MACN-CONICET) and Lic. Leonardo Silvestri (CICYTTP-CONICET-ER-UADER) are both acknowledged for processing the samples. Dr. Pedro R. Gutierrez (MACN-CONICET) is thanked for critical reading of the manuscript. We are grateful to the editor and an anonymous reviewer for their suggestions which greatly improved the manuscript. This study was supported with funds from the CONICET [grant numbers PIP 11220120100182CO, PIP 11220150100812CO].

Appendix

Complete list of species, in major groups and alphabetical order, from the studied sections with illustration reference if applicable and the occurrence in the corresponding sections. Species marked with asterisks are first records in Argentina.

Acritarchs

Ammonidium loriferum (Deunff) Hashemi and Playford, 1998 –
Ammonidium garrasinoi Ottone, 1996 (morphon) Di Pasquo et al. 2017
 (Plate I, 1) Occurrence: Angosto de San Ignacio; Angosto del Pescado.

Ammonidium spp. Occurrence: Angosto de San Ignacio, Arasayal, Balapuca.

**Baltisphaeridium juliae* Cramer, 1964 (Plate I, 2). Occurrence: Angosto del Pescado.

Bimerga bensonii Wood 1995. Occurrence: Balapuca.

Chomotriletes bistchoensis Staplin, 1961 (Plate I, 3). Occurrence: Angosto del Pescado.

Chomotriletes vedugensis Naumova, 1953 (Plate I, 4). Occurrence: Angosto del Pescado, Arasayal.

Crassiangulina sp. 1 (Plate I, 5). Occurrence: Angosto del Pescado.

Dixellophasis remota (Deunff) Playford, 1977. Occurrence: Angosto de San Ignacio, Angosto del Pescado, Balapuca.

Dixellophasis simplex Wicander and Wood, 1981. Occurrence: Angosto de San Ignacio, Angosto del Pescado.

Dixellophasis spp. Occurrence: Arasayal.

Elektoriskos sp. 1 (Plate I, 6). Occurrence: Angosto del Pescado.

Estiastra culcita Wicander, 1974. Occurrence: Arasayal.

Estiastra sp. cf. *E. culcita* Wicander, 1974 (Plate I, 7). Occurrence: Angosto del Pescado.

Estiastra sp. cf. *E. rhytidosa* Wicander and Wood, 1981 (Plate I, 8). Occurrence: Angosto del Pescado.

Estiastra spp. Occurrence: Angosto de San Ignacio.

Eupoikilofusa spp. Occurrence: Arasayal.

Evittia sp. cf. *E. sommeri* Brito 1967 (Plate I, 9). Occurrence: Angosto del Pescado.

Exochoderma arca Wicander and Wood, 1981. Occurrence: Angosto de San Ignacio, Angosto del Pescado, Balapuca.

Exochoderma irregulare Wicander, 1974. Occurrence: Balapuca.

Exochoderma triangulata Wicander and Wood, 1981. Occurrence: Angosto del Pescado, Balapuca.

Gorgonisphaeridium discissum Playford in Playford and Dring, 1981. Occurrence: Angosto de San Ignacio, Angosto del Pescado, Arasayal.

**Gorgonisphaeridium furcillatum* Wicander and Playford, 1985 (Plate I, 10). Occurrence: Angosto de San Ignacio.

Gorgonisphaeridium impexus sp. nov. (Plate I, 11–13). Occurrence: Angosto de San Ignacio.

Gorgonisphaeridium sp. cf. *G. absitum* Wicander, 1974 (Plate I, 14). Occurrence: Angosto de San Ignacio.

Gorgonisphaeridium sp. 1 (Plate I, 15). Occurrence: Angosto del Pescado.

Gorgonisphaeridium sp. 2 (Plate I, 16). Occurrence: Angosto del Pescado, Arasayal.

Hemiruptia legaultii Ottone, 1996. Occurrence: Angosto del Pescado.

Leiosphaeridia spp. Occurrence: Balapuca.

Maranhites mosesii (Sommer) Brito emend. González, 2009. Occurrence: Angosto del Pescado.

**Michrystridium flexibile* Wicander, 1974 (Plate I, 17). Occurrence: Angosto del Pescado, Arasayal.

Multiplicisphaeridium ramusculosum Deflandre emend. Lister, 1970. Occurrence: Angosto de San Ignacio, Angosto del Pescado, Balapuca.

Multiplicisphaeridium spp. Occurrence: Angosto del Pescado.

Navifusa bacilla (Deunff) Playford, 1977. Occurrence: Angosto del Pescado, Balapuca.

Neovervhachium spp. Occurrence: Arasayal.

**Ozotobrachion furcillatus* (Deunff) Playford, 1977 (Plate I, 18). Occurrence: Angosto del Pescado.

Palacanthus ledanoisii Deunff emend. Playford, 1977. Occurrence: Angosto del Pescado.

Polygonium barredae Ottone, 1996. Occurrence: Angosto del Pescado.

Polygonium spp. Occurrence: Angosto del Pescado.

Pseudolunulidia laevigata Brito and Quadros, 1985. Occurrence: Angosto del Pescado.

Sp. B in Noetinger and Di Pasquo, 2011 (Plate I, 19). Occurrence: Angosto del Pescado, Arasayal.

Stellinum micropolygonale (Stockmans and Willière) Playford, 1977. Occurrence: Angosto del Pescado.

**Stellinum oppidum* (Deunff) Le Hérisse and Deunff, 1988 in Quadros 1999 (Plate I, 20). Occurrence: Balapuca.

Stellinum spp. Occurrence: Angosto del Pescado.

Triangulina alargada Cramer, 1964. Occurrence: Angosto de San Ignacio, Angosto del Pescado, Balapuca.

Tunisphaeridium caudatum Deunff and Evitt, 1968. Occurrence: Angosto del Pescado.

Tunisphaeridium tentaculaferum (Martin) Cramer, 1971. Occurrence: Angosto del Pescado.

Tunisphaeridium spp. Occurrence: Angosto del Pescado, Balapuca.

Umbellasphaeridium sp. cf. *U. deflandrei* (Moreau-Benoit) Jardiné et al., 1972 (Plate II, 1). Occurrence: Balapuca.

Veryhachium downiei Stockmans and Willière, 1962. Occurrence: Balapuca.

**Veryhachium europaeum* Stockmans and Willière, 1960 (Plate II, 2). Occurrence: Angosto de San Ignacio.

Veryhachium lairdii Group Servais, Vecoli, Li, Molyneux, Raevskaya and Rubinstein, 2007. Occurrence: Angosto del Pescado.

**Veryhachium pastoris* Deunff, 1966 (Plate II, 3). Occurrence: Angosto de San Ignacio, Angosto del Pescado, Arasayal, Balapuca.

Veryhachium polyaster Staplin, 1961. Occurrence: Angosto del Pescado, Balapuca.

Veryhachium trispinosum Group Servais, Vecoli, Li, Molyneux, Raevskaya and Rubinstein, 2007. Occurrence: Angosto de San Ignacio, Angosto del Pescado, Arasayal, Balapuca.

Chlorophycean algae

Arkonites bilixus Legault, 1973. Occurrence: Angosto del Pescado.

Cymatiosphaera apiaria Ottone, 1996. Occurrence: Angosto de San Ignacio, Angosto del Pescado, Arasayal, Balapuca.

Cymatiosphaera perimembrana Staplin, 1961. Occurrence: Angosto de San Ignacio.

Cymatiosphaera robusta sp. nov. (Plate II, 5–6). Occurrence: Angosto del Pescado, Arasayal, Balapuca.

**Cymatiosphaera turbinata* Wicander and Loeblich, 1977 (Plate II, 7). Occurrence: Angosto de San Ignacio.

Cymatiosphaera spp. Occurrence: Arasayal, Balapuca.

Deflandrastrum? sp. 1 (Plate II, 4). Occurrence: Angosto del Pescado.

Dictyotidium variatum Playford, 1977. Occurrence: Balapuca.

Dictyotidium spp. Occurrence: Angosto de San Ignacio.

Duvernaysphaera angelae Deunff, 1964. Occurrence: Angosto del Pescado, Balapuca.

**Duvernaysphaera radiata* Brito, 1967 (Plate II, 8). Occurrence: Arasayal, Angosto de San Ignacio.

Duvernaysphaera tenuicingulata Staplin, 1961. Occurrence: Angosto del Pescado, Balapuca.

Duvernaysphaera spp. Occurrence: Balapuca.

Quadrисporites granulatus (Cramer) Cramer and Diez, 1972. Occurrence: Angosto del Pescado, Balapuca.

Quadrисporites horridus Hennelly ex Potonié and Lele, 1961. Occurrence: Balapuca.

Quadrисporites spp. Occurrence: Angosto del Pescado.

Quadrисporites variabilis (Cramer) Ottone and Rosello, 1996. Occurrence: Angosto del Pescado, Balapuca.

Leiosphaeridia spp. Occurrence: Angosto de San Ignacio, Angosto del Pescado, Arasayal, Balapuca.

Muraticavea munificus Wicander and Wood, 1981. Occurrence: Angosto del Pescado.

Polyedryxium ambitum Wicander and Wood, 1981. Occurrence: Angosto del Pescado.

Polyedryxium evolutum Deunff, 1955. Occurrence: Arasayal.

Polyedryxium fragosulum Playford, 1977. Occurrence: Angosto del Pescado.

Polyedryxium pharaonis (Deunff) Deunff, 1961. Occurrence: Angosto del Pescado, Balapuca.

Polyedryxium spp. Occurrence: Angosto del Pescado, Balapuca.
Pterospermella capitana Wicander, 1974. Occurrence: Angosto del Pescado.
Pterospermella pernambucensis (Brito) Eisenack, Cramer and Diez Rodriguez, 1973. Occurrence: Angosto del Pescado.
Pterospermella simplex sp. nov. (Plate II, 9–10). Occurrence: Angosto de San Ignacio, Angosto del Pescado, Arasayal.
Pterospermella sp. 1 (Plate II, 11). Occurrence: Angosto del Pescado.
Pterospermella spp. Occurrence: Balapuca.
Tasmanites spp. Occurrence: Angosto del Pescado.

Cryptospores

Artemopyra recticosta Breuer et al., 2007. Occurrence: Angosto del Pescado.
Cymbohilates rubinsteinae Noetinger et al., 2016(Plate II, 13). Occurrence: Angosto de San Ignacio, Arasayal.
Cymbohilates sp. 1 (Plate II, 12). Occurrence: Angosto del Pescado.
Cymbohilates sp. 2 (Plate II, 14) Occurrence: Angosto del Pescado.
Dyadospora murusattenuata morphon Strother and Traverse sensu Steemans Le Hérisson and Bozdogan, 1996. Occurrence: Angosto del Pescado.
Gneudaspora divellomedia (Tchibrikova) Balme, 1988. Occurrence: Angosto del Pescado.
**Rimosotetras problematica* Burgess, 1991 (Plate II, 15). Occurrence: Angosto del Pescado.
Tetrahedraletes medinensis Ströther and Traverse, 1979. Occurrence: Balapuca.

Spores

Acinosporites acanthomammillatus Richardson, 1965. Occurrence: Angosto del Pescado, Balapuca.
Acinosporites apiculatus (Streel) Streel, 1967. Occurrence: Angosto del Pescado, Balapuca.
Acinosporites hirsutus McGregor and Camfield, 1982. Occurrence: Angosto del Pescado.
Acinosporites ledundae Ottone, 1996. Occurrence: Angosto del Pescado.
Acinosporites lindlarensis Riegel, 1968. Occurrence: Angosto del Pescado, Balapuca.
Acinosporites macrospinosis Richardson, 1965. Occurrence: Angosto del Pescado, Balapuca.
Acinosporites spp. Occurrence: Arasayal.
Ambitisporites avitus Hoffmeister, 1959. Occurrence: Angosto del Pescado, Balapuca.
**Anapiculatisporites petilus* Richardson emend. McGregor and Camfield, 1982 (Plate II, 16). Occurrence: Angosto del Pescado, Balapuca.
Apiculatasporites microconus (Richardson) McGregor and Camfield, 1982. Occurrence: Balapuca.
Apiculatasporites ruptus sp. nov. (Plate II, 17–18). Occurrence: Angosto del Pescado, Balapuca, Angosto de San Ignacio.
Apiculatisporites grandis Menéndez and Pôthe de Baldis, 1967. Occurrence: Balapuca.
Apiculiretispora brandtii Streel, 1964. Occurrence: Angosto del Pescado, Balapuca.
Apiculiretispora densiconata Tiwari and Schaarschmidt, 1975. Occurrence: Balapuca.
Apiculiretispora laxa Amenábar, di Pasquo, Carrizo and Azcuy, 2006. Occurrence: Balapuca.
Apiculiretispora plicata (Allen) Streel, 1967. Occurrence: Angosto del Pescado, Balapuca.
Archaeozonotriletes chulus (Cramer) Richardson and Lister, 1969. Occurrence: Angosto del Pescado, Balapuca.

Archaeozonotriletes sp. 1 (Plate II, 19). Occurrence: Angosto de San Ignacio, Angosto del Pescado, Balapuca.
Archaeozonotriletes spp. Occurrence: Balapuca.
Aulicosporites spp. Occurrence: Balapuca.
Auroraspora minuta Richardson, 1965. Occurrence: Angosto del Pescado.
Biharisporites parviornatus Richardson, 1965. Occurrence: Angosto del Pescado, Balapuca.
Biornatispore sp. cf. *B. dubia* (McGregor) Steemans, 1989 (Plate III, 1). Occurrence: Angosto del Pescado.
Camarozonotriletes? *concaurus* Loboziak and Streel, 1989. Occurrence: Angosto del Pescado, Balapuca.
**Camarozonotriletes rugulosus* Breuer, Al-Ghazi, Al-Ruwaili, Higgs, Steemans and Wellman, 2007 (Plate III, 2). Occurrence: Balapuca.
**Chelinospora concinna* Allen, 1965 (Plate III, 3). Occurrence: Balapuca.
**Chelinospora ligurata* Allen, 1965 (Plate III, 4). Occurrence: Balapuca.
Chelinospora timanica (Naumova) Loboziak and Streel, 1989. Occurrence: Balapuca.
Chelinospora spp. Occurrence: Angosto de San Ignacio, Balapuca.
Convolutispora subtilis Owens, 1971. Occurrence: Angosto del Pescado, Balapuca.
**Convolutispora tegula* Allen, 1965 (Plate III, 5). Occurrence: Balapuca.
Convolutispora spp. Occurrence: Balapuca.
Corystisporites multispinosus Richardson, 1965. Occurrence: Angosto del Pescado, Balapuca.
**Corystisporites* sp. cf. *Acanthotriletes horridus* in McGregor and Camfield, 1982 (Plate III, 6). Occurrence: Balapuca.
Corystisporites undulatus Turnau, 1996. Occurrence: Angosto del Pescado.
**Craspedispora paranaensis* Loboziak, Streel and Burjack, 1988 (Plate III, 7). Occurrence: Balapuca.
Craspedispora spp. Occurrence: Balapuca.
Cymbosporites catillus Allen, 1965. Occurrence: Angosto de San Ignacio, Angosto del Pescado, Balapuca.
**Cymbosporites variegatus* Breuer and Steemans, 2013 (Plate III, 8). Occurrence: Angosto del Pescado.
Cymbosporites spp. Occurrence: Balapuca.
Densosporites devonicus Richardson, 1960. Occurrence: Angosto del Pescado.
Densosporites spp. Occurrence: Balapuca.
Dibolisporites bullatus (Allen) Riegel, 1973. Occurrence: Angosto del Pescado, Arasayal.
Dibolisporites echinaceus (Eisenack) Richardson, 1965. Occurrence: Angosto del Pescado, Balapuca.
Dibolisporites eifeliensis (Lanninger) McGregor, 1973. Occurrence: Angosto del Pescado, Balapuca.
Dibolisporites farraginis McGregor and Camfield, 1982. Occurrence: Angosto del Pescado, Balapuca.
Dibolisporites gaspiensis (McGregor) Breuer and Steemans, 2013. Occurrence: Angosto del Pescado, Balapuca.
Dibolisporites quebecensis McGregor, 1973. Occurrence: Angosto del Pescado, Balapuca.
Dibolisporites sp. cf. *D. uncatus* (Naumova) McGregor and Camfield, 1982 (Plate III, 9). Occurrence: Angosto del Pescado.
Dibolisporites sp. 1 (Plate III, 10). Occurrence: Angosto del Pescado.
**Dibolisporites* sp. 1 in Breuer and Steemans, 2013 (Plate III, 11). Occurrence: Angosto del Pescado.
**Dibolisporites tuberculatus* Breuer and Steemans, 2013 (Plate III, 12). Occurrence: Angosto del Pescado.
**Dibolisporites turriculatus* Balme, 1988 (Plate III, 13). Occurrence: Angosto de San Ignacio, Angosto del Pescado.
Dibolisporites uncatus (Naumova) McGregor and Camfield, 1982. Occurrence: Angosto del Pescado, Balapuca.

- Dibolisporites varius* Tiwari and Schaardschmidt, 1975. Occurrence: Balapuca.
- Dibolisporites* spp. Occurrence: Angosto del Pescado, Balapuca.
- Emphanisporites annulatus* McGregor, 1961. Occurrence: Angosto del Pescado, Balapuca.
- Emphanisporites epicautus* Richardson and Lister, 1969. Occurrence: Angosto del Pescado, Balapuca.
- **Emphanisporites robustus* McGregor, 1961 (Plate III, 14). Occurrence: Angosto del Pescado.
- Emphanisporites rotatus* McGregor emend. McGregor, 1973. Occurrence: Angosto del Pescado, Balapuca.
- Emphanisporites* spp. Occurrence: Arasayal.
- Emphanisporites spinaeformis* Schultz, 1968. Occurrence: Angosto del Pescado.
- **Endoculeospora altobellii* di Pasquo, 2007 [(Plate III, 15) Occurrence: Balapuca.]
- **Endosporites longiradiatus* Menéndez and Pöthe de Baldis, 1967 (Plate III, 16) Occurrence: Balapuca.
- **Geminospora antaxios* (Chibrikova) Owens, 1971 (Plate III, 17). Occurrence: Angosto del Pescado.
- Geminospora lemurata* (Balme) emend. Playford, 1983. Occurrence: Angosto del Pescado, Balapuca.
- Geminospora punctata* Owens 1971. Occurrence: Angosto del Pescado.
- Geminospora* sp. in Noetinger, 2015. Occurrence: Angosto del Pescado.
- Geminospora* sp. 1 (Plate III, 18) Occurrence: Angosto del Pescado.
- Geminospora svalbardiae* (Vigran) Allen, 1965. Occurrence: Angosto del Pescado.
- Grandispora brevizonata* (Menéndez and Pöthe de Baldis) di Pasquo, 2005. Occurrence: Angosto del Pescado, Balapuca.
- Grandispora cristata* (Menéndez and Pöthe de Baldis) Noetinger, 2015. Occurrence: Angosto del Pescado.
- Grandispora delicata* Noetinger, 2015. Occurrence: Angosto del Pescado.
- Grandispora douglastownensis* Loboziak, Streel and Burjack, 1989. Occurrence: Angosto del Pescado.
- Grandispora inculta* Allen, 1965. Occurrence: Angosto del Pescado, Balapuca.
- Grandispora mammillata* Owens, 1971. Occurrence: Balapuca.
- **Grandispora megaformis* (Richardson) McGregor, 1973 (Plate III, 19). Occurrence: Balapuca.
- Grandispora permulta* (Daemon) Loboziak, Streel and Melo, 1999. Occurrence: Angosto del Pescado.
- Grandispora protea* (Naumova) Moreau-Benoît, 1980. Occurrence: Angosto del Pescado, Balapuca.
- Grandispora pseudoreticulata* (Menéndez and Pöthe de Baldis) Ottone, 1996. Occurrence: Angosto del Pescado, Balapuca.
- Grandispora velata* (Richardson) McGregor, 1973. Occurrence: Balapuca.
- Grandispora* spp. Occurrence: Balapuca.
- Granulatisporites muninensis* Allen, 1965. Occurrence: Balapuca.
- **Granulatisporites concavus* Breuer and Steemans, 2013 (Plate III, 20). Occurrence: Angosto del Pescado.
- Leiotriletes balapucencis* di Pasquo, 2007. Occurrence: Angosto del Pescado, Balapuca.
- Leiotriletes pagius* Allen, 1965. Occurrence: Angosto del Pescado, Balapuca.
- Leiotriletes* spp. Occurrence: Angosto del Pescado, Arasayal.
- Lophotriletes devonicus* (Naumova ex Chivrikova) McGregor and Camfield, 1982. Occurrence: Angosto del Pescado, Balapuca.
- **Pero trilites heterocorpus* Steemans 1989 in Steemans, 1995 (Plate IV, 1). Occurrence: Angosto del Pescado.
- **Punctatisporites piciubaensis* Menéndez and Pöthe de Baldis, 1967 (Plate IV, 2). Occurrence: Balapuca.
- Punctatisporites* spp. Occurrence: Angosto del Pescado.
- Retusotriletes albarinii* di Pasquo and Noetinger, 2008. Occurrence: Angosto del Pescado.
- Retusotriletes maculatus* McGregor and Camfield, 1976. Occurrence: Angosto del Pescado.
- Retusotriletes ottonei* Noetinger and di Pasquo, 2011. Occurrence: Angosto del Pescado, Balapuca.
- Retusotriletes paraguayensis* Menéndez and Pöthe de Baldis, 1967. Occurrence: Angosto del Pescado, Balapuca.
- Retusotriletes simplex* Naumova, 1953. Occurrence: Angosto de San Ignacio, Angosto del Pescado.
- Retusotriletes* spp. Occurrence: Angosto de San Ignacio, Angosto del Pescado, Balapuca.
- Rhabdosporites langii* (Eisenack) Richardson, 1960. Occurrence: Angosto del Pescado.
- **Samarisporites angulatus* (Tiwari and Schaarschmidt) Loboziak and Streel, 1989 (Plate IV, 3). Occurrence: Angosto del Pescado.
- **Samarisporites praetervisus* (Naumova) Allen, 1965 (Plate IV, 4). Occurrence: Balapuca.
- Samarisporites* spp. Occurrence: Angosto del Pescado, Balapuca.
- Verruciretusispora dubia* (Eisenack) Richardson and Rasul, 1978. Occurrence: Angosto del Pescado.
- Verruciretusispora ornata* (Menéndez and Pöthe de Baldis) Pérez Leyton ex di Pasquo, 2005. Occurrence: Angosto del Pescado, Balapuca.
- Verruciretusispora* sp. 1 (Plate IV, 5) Occurrence: Angosto del Pescado.
- **Verrucosisporites onustus* Breuer and Steemans, 2013 (Plate IV, 6). Occurrence: Angosto del Pescado.
- Verrucosisporites scurrus* (Naumova) McGregor and Camfield, 1982. Occurrence: Angosto del Pescado, Balapuca.
- **Verrucosisporites* sp. 1 in Breuer and Steemans, 2013 (Plate IV, 7) Occurrence: Angosto del Pescado.
- Verrucosisporites* spp. Occurrence: Angosto del Pescado.
- Verrucosisporites tumultus* Clayton and Graham, 1974 (Plate IV, 8). Occurrence: Angosto del Pescado.

Chitinozoans

- **Ancyrochitina morzadeci* Paris, 1981 (Plate V, 1–2). Occurrence: Angosto del Pescado.
- Ancyrochitina* spp. Occurrence: Angosto del Pescado, Balapuca.
- Ancyrochitina* sp. 1 (Plate V, 3). Occurrence: Angosto del Pescado, Balapuca.
- Angochitina plicata* sp. nov. (Plate V, 4–6). Occurrence: Angosto del Pescado, Balapuca.
- Angochitina* sp. 1 (Plate V, 7). Occurrence: Balapuca.
- Cyathochitina?* sp. 1 (Plate V, 8). Occurrence: Angosto del Pescado, Balapuca.
- Fungochitina* spp. Occurrence: Angosto del Pescado, Balapuca.
- **Lagenochitina* sp. cf. *L. praeavelinoi* Grahn and Melo, 2004 (Plate V, 9). Occurrence: Angosto del Pescado.
- **Ramochitina autasmirimense* Grahn and Melo, 2004 (Plate V, 10). Occurrence: Angosto del Pescado.
- Ramochitina* spp. Occurrence: Angosto del Pescado.
- Sagenachitina* sp. 1 (Plate V, 11). Occurrence: Angosto del Pescado.
- **Spinachitina* aff. *S. biconstricta* (Lange) Grahn and Melo, 2005 (Plate V, 12). Occurrence: Angosto del Pescado.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.revpalbo.2018.07.009>.

References

- Allen, K.C., 1965. Lower and Middle Devonian spores of North and Central Vestspitsbergen. *Palaeontology* 8, 687–748.

- Amenábar, C.R., di Pasquo, M., 2008. Nuevos aportes a la palinología, cronología y paleoambiente de la Precordillera Occidental de Argentina: formaciones El Planchón, Codo (Devónico) y El Ratón (Mississippiano). *Acta geológica illosoana* 21, 3–20.
- Amenábar, C.R., Di Pasquo, M., Carrizo, H.A., Azcuy, C.L., 2006. Palynology of the Chigua (Devonian) and Malimán (carboniferous) formations in the Volcán range, San Juan Province, Argentina. Part I. Paleomicoplankton and acavate smooth and ornamented spores. *Ameghiniana* 43, 339–375.
- Amenábar, C.R., Di Pasquo, M., Carrizo, H.A., Azcuy, C.L., 2007. Palynology of the Chigua (Devonian) and Malimán (Carboniferous) formations in the Volcán Range, San Juan Province, Argentina. Part II. Cavate, pseudosaccate and cingulizolate spores. *Ameghiniana* 44, 547–564.
- Antelo Pérez, B., 1983. Formación Pescado (Río Iruya - Salta) su edad y correlación. *Rev. la Asoc. Geológica Argentina* 38, 118–125.
- Aráoz, L., Noetinger, S., Vergel, M., Del, M., Di Pasquo, M., 2016. Bioestratigrafía, paleogeografía y paleoecología del Paleozoico de Sierra de Zenta, Cordillera Oriental Argentina. Ser. correlación geológica 32, 43–64.
- Baldis, B.A., Peralta, S.H., 1999. Silúrico y Devónico de la Precordillera de Cuyo y Bloque de San Rafael. In: Caminos, R. (Ed.), *Geología Argentina, Anales. Instituto de Geología y Recursos Minerales*, Buenos Aires, Argentina, pp. 215–238.
- Balme, B.E., 1962. Upper Devonian (Frasnian) spores from the Carnarvon Basin, Western Australia. *Palaeobotanist* 9, 1–10.
- Barreda, V., 1986. Acrítarcos Givetiano-Frasnianos de la Cuenca del Noroeste, Provincia de Salta, Argentina. *Rev. Española Micropaleontol.* 18, 229–245.
- Breuer, P., Steemans, P., 2013. Devonian Spore Assemblages from Northwestern Gondwana: Taxonomy and Biostratigraphy, Special Papers in Palaeontology. The Palaeontological Association, London.
- Breuer, P., Al-Ghazi, A., Al-Ruwaili, M., Higgs, K.T., Steemans, P., Wellman, C.H., 2007. Early to Middle Devonian miospores from northern Saudi Arabia. *Rev. Micropaleontol.* 50, 27–57.
- Brito, I.M., 1967. Silurian and Devonian Acritharchs from Maranhão Basin, Brazil. *Micropaleontology* 13, 473–482.
- Cingolani, C.A., Berry, C.M., Morel, E., Tomezzoli, R., 2003. Middle Devonian lycopsids from high southern palaeolatitudes of Gondwana (Argentina). *Geol. Mag.* 139, 641–649.
- Combaz, A., 1962. Sur un nouveau type de microplanctone cénoïdal fossile du Gothlandien de Libye, *Deflandrastrum* nov. gen. *Comptes rendus l'Academie des Sci.* 225, 1977–1979.
- Cuerda, A.J., Baldis, B.A., 1971. Silúrico-Devónico de la Argentina. *Ameghiniana* 8, 128–164.
- De Amorim Gaugris, K., Grahn, Y., 2006. New chitinozoan species from the Devonian of the Paraná Basin, South Brazil, and their biostratigraphic significance. *Ameghiniana* 43, 293–310.
- de Araujo Carvalho, M., Mendonça Filho, J.G., Menezes, T.R., 2006. Paleoenvironmental reconstruction based on palynofacies analysis of the Aptian-Albian succession of the Sergipe Basin, Northeastern Brazil. *Mar. Micropaleontol.* 59, 56–81.
- Deflandre, G., 1954. Systématique des Hystrichospores et des Hystriechospores: sur l'acceptation du genre *Cymatiosphaera* O. Wetzel. *Comptes Rendu Somm. Bull. la Société Géologique Fr.* 4, 257–259.
- Di Pasquo, M., 2002. The *Crassispora kosankei*-*Cystoptychus azcuyi* palynozone of the Upper Carboniferous Tupambí formation, Tarija Basin, northern Argentina. *Rev. Palaeobot. Palynol.* 118, 47–76.
- Di Pasquo, M., 2005. Resultados palinológicos preliminares de estratos del Devónico y Carbonífero en el perfil de Balapuca, Sur de Bolivia. *La Plata* 293–298.
- Di Pasquo, M., 2007. Asociaciones palinológicas en las formaciones Los Monos (Devónico) e Itacú (Carbonífero Inferior) en Balapuca (Cuenca Tarija), sur de Bolivia. Parte 1. Formación Los Monos. *Rev. Geológica Chile* 34, 97–137.
- Di Pasquo, M.M., Noetinger, S., 2008. First record of early Devonian (Lochkovian) flora from the Santa Rosa formation-Alarache, Southern Bolivia. *Geol. Acta* 6, 191–210.
- Di Pasquo, M., Amenábar, C.R., Noetinger, S., 2009. Middle Devonian microfloras and megafloras from western Argentina and southern Bolivia: their importance in the palaeobiogeographical and palaeoclimatic evolution of western Gondwana. *Geol. Soc. London, Spec. Publ.* 314, 193–213.
- Di Pasquo, M., Noetinger, S., Isaacson, P., Grader, G., Starck, D., Morel, E., Folnagy, H.A., 2015. Mid-late Devonian assemblages of herbaceous lycophytes from northern Argentina and Bolivia: age assessment with palynomorphs and invertebrates and paleobiogeographic importance. *J. S. Am. Earth Sci.* 63, 70–83.
- Di Pasquo, M., Grader, G.W., Warren, A., Rice, B., Isaacson, P., Doughty, P.T., 2017. Palynologic delineation of the Devonian-Carboniferous boundary, West-Central Montana, USA. *Palynology* 41:189–220. <https://doi.org/10.1080/01916122.2017.1366180>.
- Dunn, D.L., Miller, T.H., 1964. A distinctive chitinozoan from the Alpena limestone (middle Devonian) of Michigan. *J. Paleontol.* 725–728.
- Eisenack, A., 1931. Neue Mikrofossilien des baltischen Silurs. *Palaeontol. Z.* 13, 74–118.
- Eisenack, A., 1955. Neue Chitinozoen aus dem Silur des Baltikums und dem Devon der Eifel. *Senckenb. Lethaea* 36, 311–319.
- Eisenack, A., 1964. Mikrofossilien aus dem Silur Gotlands. *Neues Jahrb. für Geol. und Palaontologie, Abhandlungen* 120, 308–342.
- Eisenack, A., 1972. Beiträge zur chitinozoen forschung. *Palaeontogr. Abteilung A* 140, 117–130.
- Eisenack, A., Cramer, F.H., Diez, M.D.C.R., 1973. Katalog der fossilen Dinoflagellaten, Hystrichosphären und verwandten Mikrökossilien. Band III Acritharcha 1. Teil, E. Schweizerbartsche Verlagsbuchhandlung, Stuttgart, Germany.
- Feruglio, E., 1933. Fossili devonici della Sierra Porongal nella regione subandina dell' Argentina settentrionale. *Ann. Geol. Bologna* 2, 3–22.
- Filipiak, P., 2005. Late Devonian and Early Carboniferous acritarchs and prasinophytes from the Holy Cross Mountains (Central Poland). *Rev. Palaeobot. Palynol.* 134, 1–26.
- García-Muro, V.J., Rubinstein, C.V., Steemans, P., 2014. Palynological record of the Silurian/Devonian boundary in the Argentine Precordillera, western Gondwana. *Neues Jahrb. Geol. Palaontol. Abh.* 274, 25–42.
- Ghavidel-Syooki, M., 1995. Palynostratigraphy and palaeogeography of a Palaeozoic sequence in the Hassanaqdar area, Central Alborz range, northern Iran. *Rev. Palaeobot. Palynol.* 86, 91–109.
- Gonzalez, F., Playford, C., Moreno, C., 2005. Upper Devonian biostratigraphy of the Iberian Pyrite Belt, Southwest Spain Part One: Miospores. *Palaeontogr. Abteilung B, Paläophytologie* 273, 1–52.
- Grahn, Y., 2002. Upper Silurian and Devonian Chitinozoa from central and southern Bolivia, Central Andes. *J. S. Am. Earth Sci.* 15, 315–326.
- Grahn, Y., 2005a. Silurian and Lower Devonian chitinozoan taxonomy and biostratigraphy of the Trombetas Group, Amazonas Basin, northern Brazil. *Bull. Geosci.* 80, 245–276.
- Grahn, Y., 2005b. Devonian chitinozoan biozones of Western Gondwana. *Acta Geol. Pol.* 55, 211–227.
- Grahn, Y., Melo, J.H., 2004. Integrated Middle Devonian chitinozoan and miospore zonation of the Amazonas Basin, northern Brazil. *Rev. Micropaleontol.* 47, 71–85.
- Grahn, Y., Gutiérrez, P.R., 2001. Silurian and Middle Devonian chitinozoa from the Zapla and Santa Bárbara ranges, Tarija Basin, northwestern Argentina. *Ameghiniana* 38, 35–50.
- Grahn, Y., Melo, J.H.G., Steemans, P., 2005. Integrated chitinozoan and miospore zonation of the Serra Grande Group (Silurian-Lower Devonian), Parnaíba Basin, Northeast Brazil. *Revista Española de Micropaleontología* 37, 183–204.
- Grahn, Y., Melo, J.H.G., Loboziak, S., 2006. Integrated Middle and Late Devonian miospore and chitinozoan zonation of the Parnaíba Basin, Brazil: an update. *Rev. Bras. Paleontol.* 9, 283–294.
- Grahn, Y., Mauiller, P.M., Bergamaschi, S., Bosetti, E.P., 2013. Palynology and sequence stratigraphy of three Devonian rock units in the Apucarana Sub-basin (Paraná Basin, South Brazil): additional data and correlation. *Rev. Palaeobot. Palynol.* 198, 27–44.
- Hammer, Ø., Harper, D.A.T., 2006. Paleontological data analysis. Blackwell Pub, Malden, MA.
- Helenes, J., De Guerra, C., Vasquez, J., 1998. Palynology and Chronostratigraphy of the upper cretaceous in the subsurface of the Barinas area, Western Venezuela. AAPG Bull. 82, 1308–1328.
- Hutter, T., 1979. *Alpenachitina crameri*, a new chitinozoan from the middle Devonian of Egypt. *Palynology* 3, 23–29.
- Ibrahim, A.C., 1933. Sporenformen des Ägirhorizontes des Ruhrreviers. Technische Hochschule, Berlin.
- Illich, H., Haney, F., Mendoza, M., 1981. Geochemistry of oil from Santa Cruz Basin, Bolivia. *Am. Assoc. Pet. Geol. Bull.* 65, 2388–2402.
- Inc, SASI, 1999. *SAS/STAT*.
- Jardiné, S., Combaz, A., Magloire, L., Peniguel, G., Vachey, G., 1972. Acritharchs du Silurien terminal et du Dévonien du Sahara Algérien. *Comptes Rendus Krefeld* 295–311.
- Kemp, E.M., Balme, B.E., Helby, R., Kyle, R.A., Playford, G., Price, P.L., 1977. Carboniferous and Permian palynostratigraphy in Australia and Antarctica: a review. *BMR J. Aust. Geol. Geophys.* 2, 177–208.
- Kley, J., Monaldi, C.R., 1999. Estructura de las Sierras Subandinas y del Sistema de Santa Bárbara. In: González Bonorino, G., Omarini, R., Viramonte, J. (Eds.), *Relatorio XIV Congreso Geológico Argentino. Asociación Geológica Argentina. Geología Del Noroeste Argentino*, Salta, pp. 415–425.
- Le Hérisé, A., Rubinstein, C.V., Steemans, P., 1996. Lower Devonian Palynomorphs from the Talacasto Formation, Cerro del Fuerte Section, San Juan Precordillera, Argentina. In: Fatka, O., Servais, T. (Eds.), *Acritharcha in Praha*. Acta Universitatis Carolinae, Geologica, pp. 497–515.
- Limachi, R., Goitia, V.H., Sarmiento, D., Arispe, O., Montecinos, R., Díaz Martínez, E., Dalenz-Farjat, A., Liachenco, N., Pérez Leyton, M., Aguilera, E., 1996. Estratigrafía, geoquímica, correlaciones, ambientes sedimentarios y bioestratigrafía del Silúrico-Devónico de Bolivia. *Memorias. Tarija, Bolivia*, pp. 183–197.
- Limanino, C.O., Massabie, A., Rosello, E.A., López Gamundi, O., Page, R., Jalfin, G., 1999. El Paleozoico de Ventania, Patagonia e Islas Malvinas. In: Caminos, R. (Ed.), *Geología Argentina, Anales. Instituto de Geología y Recursos Minerales*, Buenos Aires, Argentina, pp. 319–347.
- Loboziak, S., Strel, M., 1980. Miospores in givetian to lower frasnian sediments dated by conodonts from the Boulonnais, France. *Rev. Palaeobot. Palynol.* 29, 285–299.
- Loboziak, S., Strel, M., 1995. West Gondwanan aspects of the Middle and Upper Devonian miospore zonation in North Africa and Brazil. *Rev. Palaeobot. Palynol.* 86, 147–155.
- Loeblich, A.R.J., Wicander, R., 1976. Organic-walled microplankton from the Lower Devonian Late Gedinnian Haragan and Bois d'Arc formations of Oklahoma, USA. Part I. *Palaeontogr. Abteilung B* 159, 1–39.
- Marshall, J.E.A., 1996. *Rhabdosporites langii*, *Geminospora lemurata* and *Contagisporites optimus*: an origin for heterospory within the Progymnosperms. *Rev. Palaeobot. Palynol.* 93, 159–189.
- Martin, F., 1984. Acritharchs du Frasnien supérieur et du Famennien inférieur du bord méridional du Bassin de Dinant (Ardenne Belge). *Bull. Inst. Sci. Nat. Belge* 55, 1–57.
- Marynowski, L., Filipiak, P., Pisarzowska, A., 2008. Organic geochemistry and palynofacies of the Early–Middle Frasnian transition (Late Devonian) of the Holy Cross Mountains, Southern Poland. *Palaeogeogr. Palaeoclimatol. Palaeoecol.* 269, 152–165.
- McGregor, D.C., Camfield, M., 1982. Middle Devonian miospores from the Cape de Bray, Weatherall, and Hecla Bay formations of northeastern Melville Island, Canadian Arctic. *Geol. Surv. Canada* 348, 1–105.
- Mehlqvist, K., Vajda, V., Steemans, P., 2012. Early land plant spore assemblages from the late Silurian of Skåne, Sweden. *GFF* 134, 133–144.
- Melo, J.H.G., Loboziak, S., 2003. Devonian-Early Carboniferous miospore biostratigraphy of the Amazon Basin, Northern Brazil. *Rev. Palaeobot. Palynol.* 124, 131–202.

- Menéndez, C.A., Azcuy, C.L., 1969. Microflora Carbonica de la localidad de Paganzo, provincia de La Rioja. Parte I. Ameghiniana 6, 77–97.
- Menéndez, C.A., Pöthe de Baldis, E.D., 1967. Devonian spores from Paraguay. Rev. Palaeobot. Palynol. 1:161–172. [https://doi.org/10.1016/0034-6667\(67\)90118-2](https://doi.org/10.1016/0034-6667(67)90118-2).
- Milligan, G.W., 1980. An examination of the effect of six types of error perturbation on fifteen clustering algorithms. Psychometrika 45, 325–342.
- Noetinger, S., 2010. Middle–Upper Devonian palynoflora from the Tonono x-1 borehole, Salta Province, northwestern Argentina. Ameghiniana 47, 165–184.
- Noetinger, S., 2015. Spore diversity trends in the Middle Devonian of the Chaco–Salteño Plain, northwestern Argentina. Palaeogeogr. Palaeoclimatol. Palaeoecol. 417, 151–163.
- Noetinger, S., Di Pasquo, M., 2010. Palynomorphs from Abra Límite, Zenta Range, Eastern Cordillera, Northwestern Argentina. Rev. Bras. Paleontol. 13, 13–20.
- Noetinger, S., Di Pasquo, M., 2011. Devonian palynological assemblages from the San Antonio x-1 borehole, Tarija Basin, northwestern Argentina. Geol. Acta 9, 199–216.
- Noetinger, S., Di Pasquo, M., 2013. New palynological information from the subsurface Copo, Caburé and Rincón formations (upper Lochkovian – Emsian), Salta Province, Argentina. Mem. Assoc. Australas. Palaeontol. 44, 107–121.
- Noetinger, S., Di Pasquo, M., Isaacson, P., Aceñolaza, G., Vergel, M., Del, M., 2016. Integrated study of fauna and microflora from the Early Devonian (Pragian–Emsian) of northwestern Argentina. Hist. Biol. 28, 913–929.
- Noetinger, S., Pujana, R.R., Burrieza, A., Burrieza, H.P., 2017. Use of UV-curable acrylate gels as mounting media for palynological samples. Rev. del Mus. Argentino Ciencias Nat. N.S. 19, 19–23.
- Oksanen, J., Blanchet, F.G., Kindt, R., Legendre, P., Minchin, P.R., O'Hara, R.B., Simpson, G., Solymos, P., Stevens, M.H.H., Wagner, H., 2016. Package “vegan.”
- Ottone, E.G., 1996. Devonian palynomorphs from the Los Monos Formation, Tarija Basin, Argentina. Palynology 20, 105–155.
- Owens, B., 1971. Miospores from the Middle and Early Upper Devonian rocks of the Western Queen Elizabeth Island, Arctic Archipelago. Geol. Surv. Canada Pap. 70–38, 1–157.
- Paris, F., 2006. Chitinozoans: A Fascinating and Mysterious Microfossil-Group. Université de Rennes, Rennes-cedex.
- Playford, G., 1977. Lower to Middle Devonian Acrarchs of the Moose River Basin, Ontario. Geol. Surv. Canada Bull. 279, 1–87.
- Playford, G., 1981. Late Devonian acritarchs from the Gneudna Formation in the western Carnarvon Basin, western Australia. Geobios 14, 145–171.
- Playford, G., Dring, R.S., 1981. Late Devonian Acrarchs from the Carnarvon Basin, Western Australia. Special Papers in Palaeontology. The Palaeontological Association, London.
- Punt, W., Hoen, P.P., Blackmore, S., Le Thomas, A., Others, 2007. Glossary of pollen and spore terminology. Rev. Palaeobot. Palynol. 143, 1–81.
- Quadros, L.P., 1999. Silurian–Devonian acritarch assemblages from Paraná Basin: an update and correlation with Northern Brazilian basins. In: Rodrigues, M.A.C., Pereira, E. (Eds.), Ordovician – Devonian Palynostratigraphy in Western Gondwana: Update, Problems and Perspectives., Resumos Expandidos. Faculdade de Geologia da Universidade Estadual do Rio de Janeiro, Rio de Janeiro, Brasil, pp. 105–145.
- Raup, D.M., Crick, R.E., 1979. Measurement of faunal similarity in paleontology. J. Paleontol. 53, 1213–1227.
- Richardson, J.B., 1965. Middle Old Red Sandstone spore assemblages from the Orcadian Basin North-East Scotland. Palaeontology 7, 559–605.
- Richardson, J.B., Ioannides, N., 1973. Silurian palynomorphs from the Tanzezuft and Acacus Formations, Tripolitania, North Africa. Micropaleontology 19, 257–307.
- Richardson, J.B., Lister, T.R., 1969. Upper Silurian and Lower Devonian spore assemblages from the Welsh Borderland and South Wales. Palaeontology 12, 201–252.
- Richardson, J.B., McGregor, D.C., 1986. Silurian and Devonian Spore Zones of the Old Red Sandstone Continent and Adjacent Regions. Geological Survey of Canada, Ottawa, Canada.
- Richardson, J.B., Rasul, S.M., 1978. Palynomorphs in the Lower Devonian sediments from the Apley Barn Borehole, southern England. Pollen Spores 20, 423–462.
- Riegel, W., 1973. Sporenformen aus dem Heisdorf-Lauch- und nohn-schichten (Emsium und Eifelium) der Eifel, Rheinland. Palaeontogr. Abteilung B 142, 78–104.
- Rubinstein, C.V., Steemans, P., 2007. New palynological data from the Devonian Villavicencio Formation, PreCORDILLERA of Mendoza, Argentina. Ameghiniana 44, 3–9.
- Schneider, C.A., Rasband, W.S., Eliceiri, K.W., 2012. NIH Image to Image]: 25 years of image analysis. Nat. Methods 9, 671–675.
- Shi, G.R., 1993. Multivariate data analysis in palaeoecology and palaeobiogeography—a review. Palaeogeogr. Palaeoclimatol. Palaeoecol. 105, 199–234.
- Smith, A.H.V., Butterworth, M.A., 1967. Miospores in the Coal Seams of the Carboniferous of Great Britain. Papers in Palaeontology, Special.
- Staplin, F.L., 1961. Reef-controlled distribution of Devonian microplankton in Alberta. Palaeontology 4, 392–424.
- Starck, D., 1999. Los sistemas petroleros de la Cuenca Tarija. Actas. Mar del Plata, Argentina, pp. 63–82.
- Starck, D., Gallardo, E., Schulz, A., 1993. The pre-Carboniferous unconformity in the Argentine portion of the Tarija Basin. Comptes Rendus XII ICC-P. Buenos Aires, Argentina 373–384.
- Steemans, P., 1989. Palynostratigraphie de l’Eodévonien dans l’ouest de l’Europe. Prof. Pap. Mémoires Explor. Cart. Géol. Min. Belg. 27, 1–453.
- Steemans, P., 1995. Silurian and lower Emsian spores in Saudi Arabia. Rev. Palaeobot. Palynol. 89, 91–104.
- Steemans, P., Hérissé, A., Bozdogan, N., 1996. Ordovician and Silurian cryptospores and miospores from southeastern Turkey. Review of Palaeobotany and Palynology 93, 35–76.
- Strelc, M., Higgs, K., Loboziak, S., Riegel, W., Steemans, P., 1987. Spore stratigraphy and correlation with faunas and floras in the type marine Devonian of the Ardennes–Rhenish regions. Rev. Palaeobot. Palynol. 50, 211–229.
- Suárez Soruco, R., Lobo Boneta, J., 1983. La fase compresiva Eohercínica en el sector oriental de la Cuenca Cordillerana de Bolivia. Rev. Técnica YPF 9, 189–202.
- Team, R.D.C., 2013. R: A Language and Environment for Statistical Computing.
- Traverse, A., 2007. Paleopalynology. 2nd ed. Springer, Pennsylvania, USA.
- Turnau, E., Racki, G., 1999. Givetian palynostratigraphy and palynofacies: new data from the Bodzentyn Syncline (Holy Cross Mountains, Central Poland). Rev. Palaeobot. Palynol. 106:237–271. [https://doi.org/10.1016/S0034-6667\(99\)00011-1](https://doi.org/10.1016/S0034-6667(99)00011-1).
- Uriz, N.J., Cingolani, C.A., Basei, M.A.S., Blanco, G., Abre, P., Portillo, N.S., Siccardi, A., 2016. Provenance and paleogeography of the Devonian Durazno Group, southern Paraná Basin in Uruguay. J. S. Am. Earth Sci. 66, 248–267.
- Vavrdová, M., Bek, J., Dufka, P., Isaacson, P., 1996. Palynology of the Devonian (Lochkovian to Tournaisian) sequence, Madre de Dios Basin, northern Bolivia. Bull. Czech Geol. Surv. 71, 333–350.
- Volkheimer, W., Mendoza, D., Salas, A., 1986. Devonian chitinozoans from northwestern Argentina. Palaeontogr. Abteilung B 173, 229–251.
- Wauthoz, B., Dorning, K.J., Le Hérissé, A., 2003. *Crassiangulina variacornuta* sp. nov. from the late Llandovery and its bearing on Silurian and Devonian acritarch taxonomy. Bull. la Société géologique Fr. 174, 67–81.
- Wellman, C.H., Habgood, K., Jenkins, G., Richardson, J.B., 2000. A new plant assemblage (microfossil and megafossil) from the Lower Old Red Sandstone of the Anglo-Welsh Basin: its implications for the palaeoecology of early terrestrial ecosystems. Rev. Palaeobot. Palynol. 109, 161–196.
- Wetzel, O., 1933. Die in organischer Substanz erhaltenen Mikrofossilien des baltischen Kreide-Feuersteins mit einem sedimentpetrographischen und stratigraphischen Anhang (Schlub). Palaeontogr. Abteilung A 78, 1–110.
- Wicander, R., 1974. Upper Devonian-Lower Mississippian Acrarchs and Prasinophycean Algae from Ohio, USA. Palaeontogr. Abteilung B 148, 9–43.
- Wicander, R., Loeblich, A.R.J., 1977. Organic-walled microphytoplankton and its stratigraphic significance from the Upper Devonian Antrim Shale, Indiana, USA. Palaeontogr. Abteilung B 160, 129–165.
- Wicander, R., Playford, G., 1985. Acrarchs and spores from the Upper Devonian Lime Creek Formation, Iowa, USA. Micropaleontology 97–138.
- Wicander, R., Playford, G., 2013. Marine and terrestrial Palynoforas from transitional Devonian-Mississippian strata, Illinois Basin, USA. Bol. Geol. y Min. 124, 589–637.
- Wicander, R., Wood, G.D., 1981. Systematics and biostratigraphy of the organic-walled microphytoplankton from the Middle Devonian (Givetian) Silica Formation, Ohio, USA. AAPG Contributions. American Association of Stratigraphic Palynologists Foundation.